BIOLOGY LETTERS

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Cite this article: Chalkowski K, Wilson AE, Lepczyk CA, Zohdy S. 2019 Who let the cats out? A global meta-analysis on risk of parasitic infection in indoor versus outdoor domestic cats (*Felis catus*). *Biol. Lett.* **15**: 20180840. http://dx.doi.org/10.1098/rsbl.2018.0840

Received: 29 November 2018 Accepted: 25 March 2019

Subject Areas:

ecology, health and disease and epidemiology

Keywords:

felid, latitude, pathogen, pet, transmission, zoonotic

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Electronic supplementary material is available online at https://dx.doi.org/10.6084/m9. figshare.c.4462778.



Pathogen biology

Who let the cats out? A global meta-analysis on risk of parasitic infection in indoor versus outdoor domestic cats (*Felis catus*)

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Parasitic infection risks in domestic animals may increase as a result of outdoor activities, often leading to transmission events to and from owners, other domestic animals and wildlife. Furthermore, outdoor access has not been quantified in domestic animals as a risk factor with respect to latitude or parasite transmission pathway. Cats are an ideal model to test parasitic infection risk in outdoor animals because there have been many studies analysing this risk factor in this species; and there is a useful dichotomy in cat ownership between indoor-only cats and those with outdoor access. Thus, we used meta-analysis to determine whether outdoor access is a significant risk factor for parasitic infection in domestic pet cats across 19 different pathogens including many relevant to human, domestic animal and wildlife health, such as Toxoplasma gondii and Toxocara cati. Cats with outdoor access were 2.77 times more likely to be infected with parasites than indoor-only cats. Furthermore, absolute latitude trended towards significance such that each degree increase in absolute latitude increased infection likelihood by 4%. Thus, restricting outdoor access can reduce the risk of parasitic infection in cats and reduce the risk of zoonotic parasite transmission, spillover to sympatric wildlife and negative impacts on feline health.

1. Background

Domestic animals, including pets, are responsible for spreading pathogens to humans and sympatric wildlife [1–3]. Notable examples include dogs transmitting rabies to humans [4] or cattle transmitting *Cryptosporidium paroum* to humans and sympatric wild ruminants [5,6]. However, relatively few domestic animals have such stark dichotomies regarding outdoor access, where environmental contact can, therefore, be evaluated as a means of exposure. Understanding how outdoor access affects infection, and infection by which pathogens are most affected by this risk factor, can have important implications when mitigating parasite transmission among domestic animals, humans and wildlife.

A model organism that is widespread and lives in close proximity to humans is the domestic cat (*Felis catus*), which has coexisted with humans globally for millennia (*ca* 9500 years; [7,8]). In fact, pet cats often sit on their owners' laps and sleep in their beds [9]. Furthermore, cats are common as pets around the world, with an estimated 89–90 million in the USA alone [10]. Given that cats are widespread and associated with humans, risk factors for parasitic infections in pet cats are important for zoonotic parasite transmission, with implications for cat health as well as spillover of parasites to sympatric wildlife [11,12].

Table 1. Host ranges of pathogens analysed in this study.

pathogen	hosts	citation(s
Aelurostrongylus abstrusus	caracal (Caracal caracal), lion (Panthera leo), serval (Leptailurus serval)	[21]
Cystoisospora felis	Felidae (including European wild cat (<i>Felis sylvestris</i>), ocelot (<i>Felis pardalis</i>), serval (<i>Felis serval</i>), tiger (<i>Leo tigris</i>), jaguar (<i>Leo onca</i>), Eurasian lynx (<i>Lynx lynx</i>)), house mouse (<i>Mus musculus</i>), golden hamster (<i>Mesocricetus auratus</i>)	
Cystoisospora revolta	Felidae (including European wild cat, jungle cat (<i>Felis chaus</i>), tiger, leopard (<i>Leo pardus</i>)), house mouse, opossum (<i>Didelphis virginiana</i>), Norway rat (<i>Rattus norvegicus</i>), golden hamster	
<i>Cytauxzoon</i> spp.	meerkat (<i>Suricata suricatta</i>), bobcat (<i>Lynx rufus</i>), cougar (<i>Puma concolor</i>), Florida panther (<i>Felix concolor coryi</i>), ocelot, puma (<i>Puma yagouaroundi</i>), jaguar (<i>Panthera onca</i>)	[24–27]
Dipylidium caninum	crab-eating fox (<i>Cerdocyon thous</i>), red fox (<i>Vulpes vulpes</i>), golden jackal (<i>Canis aureus</i>), wolf (<i>Canis lupus</i>)	[28,29]
feline coronavirus	Felidae (including cheetah (Acinonyx jubatus), European wildcat, Canada lynx (Lynx canidensis))	[30-32]
feline leukemia virus	Felidae (including European wildcat), spotted hyena (Crocuta crocuta)	[31-33]
feline immunodeficiency virus	Felidae (including European wildcat, sand cat (Felis margarita)), spotted hyena	[31-33]
Giardia lamblia	<i>Giardia</i> affects a large number of mammal and bird species, but it appears that the assemblage in domestic cats is not found in other species	[34]
Hemoplasma spp.	Iberian lynx, Eurasian lynx, European wildcat, lion, puma, oncilla (<i>Leopardus tigrinis</i>), Geoffroy's cat (<i>Leopardus geoffroyi</i>), margay (<i>Leopardus wiedii</i>), ocelot	[35]
Hepatozoon spp.	coyote (<i>Canis latrans</i>), bobcat, ocelot	[36]
<i>Mycoplasma</i> spp.	Iberian lynx, Eurasian lynx, lion, European wildcat	[37]
Neospora caninum	Canidae (including red fox, grey fox (<i>Urocyon cinereoargeneteus</i>), Australian dingo (<i>Canis familiaris dingo</i>), Chiloé fox (<i>Pseudolapex fulvipes</i>)), cheetah, raccoon (<i>Procyon lotor</i>)	[38,39]
<i>Taenia</i> spp.	several Taenia species infect a wide variety of carnivores	[40]
Toxocara cati	can infect small mammals (including Guinea pigs (<i>Cavia porcellus</i>) and house mouse but data are lacking	[41]
Toxoplasma gondii	wide host range of almost any bird or mammal evaluated	[42]
Trichuris spp.	widespread across mammal species depending on species of Trichuris	[43,44]
Troglostrongylus brevior	European wild cat	[45]

Domestic pet cats allowed outdoors can also pose health risks to cat owners [13–19]. For instance, *Toxoplasma gondii* (the causative agent of toxoplasmosis; [15]) and *Bartonella henslae* (which causes cat-scratch disease; [17]), both infect people worldwide. In addition, there are many infectious diseases that have health consequences for cats themselves. For example, feline immunodeficiency virus (FIV) causes immunosuppression which can increase susceptibility to other infections [20]. Finally, interactions with sympatric wildlife may result in spillover of parasites from domestic cats (table 1). For example, domestic cats have been responsible for the spread of FIV to mountain lions (*Puma concolor*) and feline panleukopenia to the Florida panther (*Puma concolor coryi*) [11,12].

Many parasites known to infect cats have life cycles involving transmission from the soil, prey, or other cats [15,46–49]. Here, we hypothesize that cats with outdoor access (freeroaming) will be more likely to be infected with parasites than indoor-only cats. To test our hypothesis, we conducted a meta-analysis of outdoor access as a risk factor for infection across 19 pathogens and 16 countries. Because differences in risk of infection may exist owing to changes in pathogen diversity (i.e. richness and abundance) across transmission type and space [50–52], we considered transmission type and latitude as separate moderators.

2. Results

(a) Overall effects

Our synthesis incorporated 21 studies with 31 sets of infection prevalence between indoor-only cats and those with outdoor access (table 2). Among the 21 studies, 19 parasites were analysed (see electronic supplementary material, figure S1 for odds ratios (OR) by parasite and study). According to the overall model, cats with outdoor access are 2.77 (95% confidence limits (95% CL) = 2.10-3.67; p < 0.0001) times as likely to be infected with parasites as indoor-only cats (figure 1). Heterogeneity, or differences in outcomes between studies [70], in the overall model was high (I^2 = 84.02%). The publication bias analysis estimated six missing studies on the left side of the funnel plot (figure 2a,b) and incorporation of these randomly created studies using the trim and fill technique still resulted in the effect of outdoor access as a significant risk factor (2.39 OR; p < 0.0001).

Table 2. Pathogen prevalence in domestic cats (*Felis catus*) in this study by country.

pathogen	country	prevalence	citation
Aelurostrongylus abstrusus	Cyprus	0.02	[53]
Cystoisospora revolta	Cyprus	0.12	[53]
<i>Cytauxzoon</i> spp.	Spain	0.01	[54]
Dipylidium caninum	Cyprus	0.01	[53]
feline coronavirus	Australia	0.41	[55]
FIV	Australia	0.10	[56]
		0.31	[57]
	Canada	0.63	[58]
Giardia lamblia	Cyprus	0.07	[53]
Hemoplasma spp.	Chile	0.15	[59]
Mycoplasma spp.	Spain	0.07	[54]
	Germany	0.10	[60]
	Switzerland	0.09	[61]
Neospora caninum	Brazil	0.03	[62]
<i>Taenia</i> spp.	Cyprus	0.01	[53]
<i>Toxocara</i> spp.	Cyprus	0.12	[53]
	Netherlands	0.05	[63]
Toxoplasma gondii	Estonia	0.62	[64]
	Pakistan	0.26	[65]
	Latvia	0.53	[66]
	Romania	0.48	[67]
Trichuris spp.	St Kitts	0.22	[68]
Troglostrongylus spp.	Cyprus	0.05	[53]
	Netherlands	0.20	[69]

(b) Moderators

Transmission type was not a significant moderator (p = 0.62; figure 1), but infection risk in indoor-only pet cats versus those with outdoor access trended towards significance with latitude (figure 2). Specifically, for every degree increase in absolute latitude, cats with outdoor access were 4% more likely to be infected with parasites (95% CL = 1.0-7.0%; p = 0.081; figure 2*a*). Heterogeneity decreased considerably with the inclusion of this moderator to $I^2 = 55.7\%$ (from 84.0%), suggesting differences in latitude may account for a significant portion of the variation among studies.

To determine the true effect of increasing latitude (since OR is only a relative comparison of indoor-only and outdoor cats), we also conducted a meta-regression using a raw proportion of the total number of infected cats, with absolute latitude as a moderator. In this model, the overall proportion of infected cats significantly increased, by 0.7% (95% CL = 0.17-1.3%; OR 95% CL = 1.01-1.07; p = 0.010) for each degree latitude increase (figure 2*b*), indicating that increasing risk of infection in cats with outdoor access with increasing latitude is an important interaction.

3. Discussion

Outdoor access is a significant risk factor for parasitic infection in pet cats, where cats with outdoor access were 2.77

times more likely to be infected with parasites than indoor-only cats, demonstrating support for our hypothesis. Of the 21 studies we included, only three suggested non-significantly higher risk of infection in indoor-only cats. Furthermore, latitude had a marginally significant effect on the likelihood of infection. While there was publication bias indicating positive results for outdoor access as a risk factor, following the trim and fill method the effects were similar and still significant, suggesting publication bias did not influence the significance of the meta-analysis results.

The parasites we analysed have relevance to zoonotic parasite transmission, feline health and wildlife conservation. Given the association between humans and domestic cats [9], habitat and lifestyle risk factors ought to be investigated with respect to zoonotic parasite infection. Furthermore, despite ubiquity of domestic cats, cat-human transmission is likely under-reported [71].

Not only are parasitic infections impactful to feline health, they are also relevant to wildlife. Parasites of domestic cats have already been reported in sympatric wild congeners, such as FIV in cougars (*Felis concolor*) and *Candidatus* Mycoplasma haemominutum in wild felids deriving from domestic cats [11,12,37]. Positive associations between feline herpesvirus type 1 (FHV-1) and *Bartonella* in cougars and urban land-use have also been reported, suggesting interactions with domestic cats [72]. However, further investigation into infection prevalence in wild populations and risk factors for transmission between domestic cats and these species is warranted [12].

Among the transmission types analysed (i.e. direct, vectorborne and environmental), none differed significantly from either of the others with respect to effect of outdoor access on parasitic infection. Two explanations are the small sample size between groups or within studies, and high variability across studies. Additionally, a Bayesian approach using a Markov chain Monte Carlo method may have better accounted for this uncertainty [73]. Directly transmitted parasites (i.e. cat–cat transmission), such as FIV, were not significantly different from other transmission types with respect to outdoor access, which suggests these parasites may be more frequently encountered through contact with feral populations or other pet cats allowed outdoor access rather than from cats in shelters or the household.

Latitude as a moderator on infection risk in cats with outdoor access trended towards a significant positive effect. The trend identified ran contrary to what has been demonstrated for parasite richness and diversity, which typically decrease with increasing latitude [50-52]. Although one might assume that higher parasite diversity results in higher infection risk in hosts, there have been multiple findings demonstrating the opposite-that infection rates decrease with higher parasite diversity [74,75]-which is consistent with our finding that cats with outdoor access in northern regions are at greater risk of infection. Interestingly, these results were also consistent with global patterns of zoonoses in rodents, a common prey of domestic cats, where higher latitudes saw greater numbers of species carrying zoonoses [76]. Higher latitudes also predicted greater risk of helminth parasites from wildlife found in domestic animals [2].

Organizations including the American Bird Conservancy (ABC) and People for the Ethical Treatment of Animals (PETA) have created campaigns that raise awareness about the detrimental impacts of cats with outdoor access in relation to feline health and impacts on wildlife [77,78],

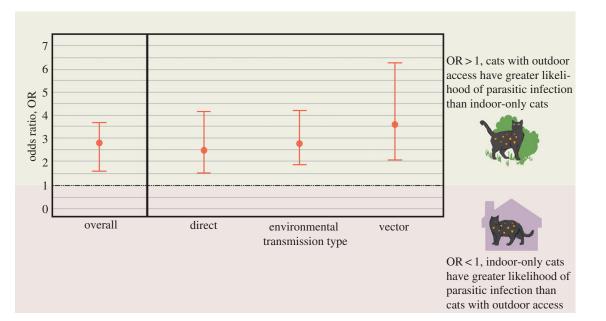


Figure 1. Overall effect size and transmission type effect sizes for infection prevalence in cats with outdoor access versus indoor-only cats. Cats with outdoor access are 2.77 (95% CL = 2.10-3.67; p < 0.0001) times as likely to be infected with parasites as indoor-only cats. Transmission types include environmental (soil-borne and intermediate hosts), vector-borne and direct. Transmission type was not a significant moderator (p = 0.62) for outdoor access on infection prevalence in domestic pet cats.

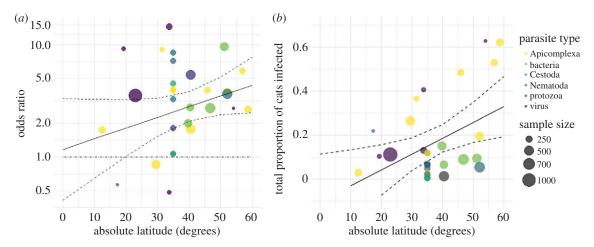


Figure 2. (*a*) The relationship between odds ratio for each study/parasite in domestic pet cats across a range of latitudes. For every degree increase in latitude, cats with outdoor access were 1.04 times as likely to be infected with parasites (95% CL = 1.01 - 1.07). Latitude as a moderator to indoor/outdoor infection risk was trending towards significance (p = 0.081). (*b*) Total proportions of infected cats for each study/parasite across a range of latitudes where overall proportion of infected cats significantly increased, by 0.7% (95% CL = 0.17 - 1.3%; p = 0.010) for each degree latitude increase.

though allowing pet cats outdoors is still common occurrence [79,80]. Increased awareness of the risks involved in outdoor access is one facet, but legislation restricting outdoor access in cats would be an ideal outcome [81]. Despite hurdles in enacting new legislation, this issue has a relatively simple solution—keep cats indoors.

Domestic cats act as potent reservoirs for parasites transmissible to wildlife and humans [82–84], and are a unique model for understanding pathogen transmission dynamics given their global ubiquity and contact with humans, other animals and the environment. Our analysis is the first to our knowledge to summarize across many parasites and geographical localities that outdoor access increases the odds of parasitic infection in pet cats as a model for domestic animals. Future research might investigate this risk factor across other domestic species and across factors, such as land use and presence of sympatric congeners. While we do not necessarily advocate that all domestic animals be restricted indoors, determining routes and risk factors of transmission with respect to environmental contact may be useful in mitigating parasitic infection in domestic animals.

4. Methods

(a) Literature search

A literature search using Web of Science was conducted on 11 January 2018, following PRISMA [85] guidelines, with the following keywords: 'feral cat' OR 'feral dog*' AND 'infect*' OR 'parasit*' OR 'disease*' OR 'virus*', excluding reviews. This search returned 500 research articles, which were manually sorted for relevance. Final output was based on the following exclusion criteria: review articles; case studies; sample size less than 20 cats; lack of comparison between indoor-only versus

outdoor access pet domestic cats; or outdoor access group included feral or stray cats.

An additional search was performed in Web of Science on 31 May 2018, using the following keywords: 'domestic cat*' OR 'pet cat*' OR 'Felis catus' AND 'outdoor access' AND TOPIC: ('infection*' OR 'parasit*' OR 'disease*' OR 'pathogen*' OR 'virus*' OR 'sick*' OR 'illness*'), which returned 213 additional articles. One search was conducted in Google Scholar using the keywords as follows: domestic OR pet cat OR Felis catus, outdoor access, infection* OR parasit*. This Google Scholar search returned 1190 results. We manually sorted through the first 100 studies using the exclusion criteria described above. After manually sorting the original output of 813 studies, 21 studies fitted the inclusion criteria and were used in the meta-analysis [86] (see https://figshare.com/s/3eebaf42e161c0e7e1ef to access dataset).

(b) Treatment of moderators

Parasite transmission type included direct, vector-borne and environmental pathways (see electronic supplementary material, figure S2 for list of citations for each parasite). Latitude of each study was determined using Google Earth by selecting the middle of the smallest geographical area provided (such as country, state/province or city). Studies that included multiple countries were removed from analysis of this moderator.

(c) Statistical analysis

All analyses were completed in R v.1.1.453 using the *metafor* package for random effects models to account for betweenstudy heterogeneity using the OR effect size [87,88], where an OR is the probability of an outcome as related to an exposure [89]. Here, the outcome is likelihood of infection as related to outdoor access as the exposure mechanism. OR = 1 means outdoor access does not affect the likelihood of infection; OR < 1 (upper 95% CI is less than 1) means outdoor access is associated with lower odds of infection; and OR > 1 (lower 95% CI is greater than 1) means outdoor access is associated with greater odds of infection. We considered p < 0.05 to indicate the significance of effect size. Two moderators, transmission type and latitude, were evaluated using mixed effects models.

To estimate heterogeneity across studies, we used l^2 , where a value of 0% indicates no heterogeneity; 25%, low heterogeneity; 50%, moderate; and 75% is considered high heterogeneity [90]. To test for publication bias, we used a trim and fill method to estimate the number of missing studies [91].

Data accessibility. Literature search: Figshare repository figshare.com/s/ 3eebaf42e161c0e7e1ef [86]. R code in analyses: Figshare repository figshare.com/s/a334c7815b128cb63b98 [87].

Authors' contributions. K.C. designed the study, conducted literature review and analyses, and wrote the manuscript; A.E.W. participated in statistical analyses, study design and manuscript writing; C.A.L. participated in statistical analyses and manuscript writing; S.Z. participated in statistical analyses and manuscript writing. All authors gave approval for the final version of this manuscript, and agree to be accountable for its content.

Competing interests. The authors declare no competing interests.

Funding. K.C. was supported by the Auburn University Cell and Molecular Biology Fellowship Program. Funding for S.Z. was provided by a Young Investigator Award from the USDA National Institute of Food and Agriculture, and CDC-RFA- CK14-1401PPHF. This project was supported by the Alabama Agricultural Experiment Station, the Hatch Program of the National Institute of Food and Agriculture, U.S. Department of Agriculture.

Acknowledgements. Thanks to the Auburn School of Forestry and Wildlife Science SQUAD (Solving Quantitative, Unusual and Awesome Dilemmas); Todd Steury and Ash Abebe for help with data analysis and interpretation of results; and Patricia Hartman for help conducting the literature search.

References

- Landaeta-Aqueveque C, Henríquez A, Cattan PE. 2014 Introduced species: domestic mammals are more significant transmitters of parasites to native mammals than are feral mammals. *Int. J. Parasitol.* 44, 243–249. (doi:10.1016/j.ijpara. 2013.12.002)
- Wells K, Gibson DI, Clark NJ, Ribas A, Morand S, McCallum HI. 2018 Global spread of helminth parasites at the human – domestic animal – wildlife interface. *Glob. Chang. Biol.* 24, 3254–3265. (doi:10.1111/gcb.14064)
- Clark NJ, Seddon JM, Šlapeta J, Wells K. 2018 Parasite spread at the domestic animal - wildlife interface: anthropogenic habitat use, phylogeny and body mass drive risk of cat and dog flea (*Ctenocephalides* spp.) infestation in wild mammals. *Parasites Vectors* **11**, 8. (doi:10.1186/s13071-017-2564-z)
- Tang X, Luo M, Zhang S, Fooks AR, Hu R, Tu C. 2005 Pivotal role of dogs in rabies transmission, China. *Emerg. Infect. Dis.* **11**, 1970 – 1972. (doi:10.3201/ eid1112.050271)
- Alves M, Xiao L, Sulaiman I, Lal AA, Matos O, Antunes F. 2003 Subgenotype analysis of *Cryptosporidium* isolates from humans, cattle, and zoo ruminants in Portugal. *J. Clin. Microbiol.* 41,

2744–2747. (doi:10.1128/JCM.41.6.2744-2747. 2003)

- Alves M, Xiao L, Antunes F, Matos O. 2006 Distribution of *Cryptosporidium* subtypes in humans and domestic and wild ruminants in Portugal. *Parasitol. Res.* 99, 287–292. (doi:10.1007/s00436-006-0164-5)
- Driscoll CA *et al.* 2007 The Near Eastern origin of cat domestication. *Science* **317**, 519–523. (doi:10. 1126/science.1139518)
- Fleming PA, Bateman PW. 2018 Novel predation opportunities in anthropogenic landscapes. *Anim. Behav.* 138, 145–155. (doi:10.1016/j.anbehav. 2018.02.011)
- Chomel BB, Sun B. 2011 Zoonoses in the bedroom. Emerg. Infect. Dis. 17, 167–172. (doi:10.3201/ eid1702.101070)
- Lepczyk CA, Duffy DC. 2018 Feral cats. In *Ecology* and management of terrestrial vertebrate invasive species in the United States (eds WC Pitt, J Beasley, GW Witmer), pp. 269–310. Boca Raton, FL: CRC Press.
- Jessup DA, Pettan KC, Lowenstine LJ, Pedersen NC. 1993 Feline leukemia virus infection and renal spirochetosis in a free-ranging cougar (*Felis concolor*). J. Zoo Wildl. Med. 24, 73–79.

- Roelke ME, Forrester DJ, Jacobson ER, Kollias GV, Scott FW, Barr MC, Evermann JF, Pirtle EC. 1993 Seroprevalence of infectious disease agents in freeranging Florida panthers (*Felis concolor coryi*). *J. Wildl. Dis.* 29, 36–49. (doi:10.7589/0090-3558-29.1.36)
- Lepczyk CA, Lohr CA, Duffy DC. 2015 A review of cat behavior in relation to disease risk and management options. *Appl. Anim. Behav. Sci.* 173, 29–39. (doi:10.1016/j.applanim.2015.07.002)
- Loyd KAT, Hernandez SM, Abernathy KJ, Shock BC, Marshall GJ. 2013 Risk behaviours exhibited by free-roaming cats in a suburban US town. *Vet. Rec.* **173**, 295. (doi:10.1136/vr. 101222)
- Hill D, Dubey JP. 2002 Toxoplasma gondii: transmission, diagnosis and prevention. Clin. Microbiol. Infect. 8, 634–640. (doi:10.1046/j.1469-0691.2002.00485.x)
- Fisher M. 2003 *Toxocara cati*: an underestimated zoonotic agent. *Trends Parasitol.* **19**, 167–170. (doi:10.1016/S1471-4922(03)00027-8)
- Chomel BB, Boulouis H-J, Maruyama S, Breitschwerdt EB. 2006 *Bartonella* spp. in pets and effect on human health. *Emerg. Infect. Dis.* 12, 389–394. (doi:10.3201/eid1203.050931)

- Luft BJ, Remington JS. 1992 Toxoplasmic encephalitis in AIDS. *Clin. Infect. Dis.* 15, 211–222. (doi:10.1093/clinids/15.2.211)
- Baliu C, Sanclemente G, Cardona M, Castel MA, Perez-Villa F, Moreno A, Cervera C. 2014 Toxoplasmic encephalitis associated with meningitis in a heart transplant recipient. *Transpl. Infect. Dis.* 16, 631–633. (doi:10.1111/tid.12242)
- Sparkes AH, Hopper CD, Millard WG, Gruffydd-Jones TJ, Harbour DA. 1993 Feline immunodeficiency virus infection clinicopathologic findings in 90 naturally occurring cases. J. Vet. Intern. Med. 7, 85–90. (doi:10.1111/j.1939-1676.1993.tb03174.x)
- Di Cesare A, Laiacona F, Iorio R, Marangi M, Menegotto A. 2016 *Aelurostrongylus abstrusus* in wild felids of South Africa. *Parasitol. Res.* **115**, 3731–3735. (doi:10.1007/s00436-016-5134-y)
- Queshi T. 2014 *Isospora felis*. American Association of Veterinary Parasitologists. See http://www.aavp. org/wiki/catprotozoa/coccidia-apicomplexan/ isospora-felis/ (accessed 16 January 2018).
- Bowman A. 2014 Isospora rivolta. American Association of Veterinary Parasitologists. See http:// www.aavp.org/wiki/catprotozoa/coccidiaapicomplexan/isospora-rivolta/ (accessed 16 January 2018).
- 24. Leclaire S, Menard S, Berry A. 2015 Molecular characterization of *Babesia* and *Cytauxzoon* species in wild South-African meerkats. *Parasitology* **142**, 543–548. (doi:10.1017/ S0031182014001504)
- Shock BC *et al.* 2011 Distribution and prevalence of *Cytauxzoon felis* in bobcats (*Lynx rufus*), the natural reservoir, and other wild felids in thirteen states. *Vet. Parasitol.* **175**, 325–330. (doi:10.1016/j.vetpar. 2010.10.009)
- Butt MT, Bowman D, Barr MC, Roelke ME. 1991 latrogenic transmission of *Cytauxzoon felis* from a Florida panther (*Felix concolor coryi*) to a domestic cat. *J. Wildl. Dis.* 27, 342–347. (doi:10.7589/0090-3558-27.2.342)
- Bowman A. 2014 Cytauxzoon felis. American Association of Veterinary Parasitologists. See http:// www.aavp.org/wiki/catprotozoa/coccidiaapicomplexan/piroplasms-cytauxzoon-babesia/ cytauxzoon-felis/ (accessed 16 January 2019).
- Dalimi A, Sattari A, Motamedi G. 2006 A study on intestinal helminthes of dogs, foxes and jackals in the western part of Iran. *Vet. Parasitol.* 142, 129–133. (doi:10.1016/j.vetpar.2006.06.024)
- Segovia JM, Torres J, Miquel J, Llaneza L, Feliu C.
 2001 Helminths in the wolf, *Canis lupus*, from north-western Spain. *J. Helminthol.* **75**, 183–192. (doi:10.1079/J0H200152)
- Heeney JL, Caro T. 1990 Prevalence and implications of feline coronavirus infections of captive and freeranging cheetahs (*Acinonyx jubatus*). J. Virol. 64, 9.
- Daniels MJ, Golder MC, Jarrett O, MacDonald DW. 1999 Feline viruses in wildcats from Scotland. *J. Wildl. Dis.* **35**, 121–124. (doi:10.7589/0090-3558-35.1.121)
- 32. Ostrowski S, Van Vuuren M, Lenain DM, Durand A. 2003 A serologic survey of wild felids from central

west Saudi Arabia. *J. Wildl. Dis.* **39**, 696-701. (doi:10.7589/0090-3558-39.3.696)

- Harrison TM, Mazet JK, Holekamp KE, Dubovi E, Engh AL, Nelson K, Van Horn RC, Munson L. 2004 Antibodies to canine and feline viruses in spotted hyenas (*Crocuta crocuta*) in the Masai Mara National Reserve. J. Wildl. Dis. 40, 1–10. (doi:10.7589/0090-3558-40.1.1)
- Feng Y, Xiao L. 2011 Zoonotic potential and molecular epidemiology of *Giardia* species and giardiasis. *Clin. Microbiol. Rev.* 24, 110–140. (doi:10.1128/CMR.00033-10)
- Willi B et al. 2007 Worldwide occurrence of feline Hemoplasma infections in wild felid species. J. Clin. Microbiol. 45, 1159–1166. (doi:10.1128/JCM. 02005-06)
- Mercer SH, Jones LP, Rappole JH, Twedt D, Laack LL, Craig TM. 1988 *Hepatozoon* sp. in wild carnivores in Texas. *J. Wildl. Dis.* 24, 574–576. (doi:10.7589/ 0090-3558-24.3.574)
- Kellner A, Carver S, Scorza V, McKee CD, Lappin M, Crooks KR, VandeWoude S, Antolin MF. 2018 Transmission pathways and spillover of an erythrocytic bacterial pathogen from domestic cats to wild felids. *Ecol. Evol.* 8, 9779–9792. (doi:10. 1002/ece3.4451)
- McAllister MM, Dubey JP, Lindsay DS, Jolley WR, Wills RA, McGuire AM. 1998 Dogs are definitive hosts of *Neospora caninum*. *Int. J. Parasitol*. 28, 1473-1478. (doi:10.1016/S0020-7519(98) 00138-6)
- Dubey JP. 2003 Review of *Neospora caninum* and neosporosis in animals. *Korean J. Parasitol.* 41, 1-16. (doi:10.3347/kjp.2003.41.1.1)
- Hoberg EP. 2006 Phylogeny of *Taenia*: species definitions and origins of human parasites. *Parasitol. Int.* 55, S23-S30. (doi:10.1016/j.parint. 2005.11.049)
- Nolan T. 2014 *Toxocara cati*. American Association of Veterinary Parasitologists. See http://www.aavp.org/ wiki/nematodes/ascaridida/toxocara-cati/ (accessed 16 January 2019).
- 42. Dubey JP. 2009 *Toxoplasmosis of animals and humans*. Boca Raton, FL: CRC Press.
- Ghai RR, Simons ND, Chapman CA, Omeja PA, Davies TJ, Ting N, Goldberg TL. 2014 Hidden population structure and cross-species transmission of whipworms (*Trichuris* sp.) in humans and nonhuman primates in Uganda. *PLoS Negl. Trop. Dis.* 8, e0003256. (doi:10.1371/journal.pntd.0003256)
- 44. Xie Y, Zhao B, Hoberg EP, Li M, Zhou X, Gu X, Lai W, Peng X, Yang G. 2018 Genetic characterisation and phylogenetic status of whipworms (*Trichuris* spp.) from captive non-human primates in China, determined by nuclear and mitochondrial sequencing. *Parasites Vectors* **11**, 5. (doi:10.1186/ s13071-018-3100-5)
- Deak G, Ionică AM, Mihalca AD, Gherman CM. 2017 *Troglostrongylus brevior*: a new parasite for Romania. *Parasites Vectors* 10, 599. (doi:10.1186/ s13071-017-2551-4)
- Beaver P. 1975 Biology of soil-transmitted helminths: the massive infection. *Health Lab. Sci.* 12, 116–125.

- Hardy WD, Old LJ, Hess PW, Essex M, Cotter S. 1973 Horizontal transmission of feline leukaemia virus. *Nature* 244, 266–269. (doi:10.1038/ 244266a0)
- Allen HA. 2015 Characterizing zoonotic disease detection in the United States: who detects zoonotic disease outbreaks & how fast are they detected? *J. Infect. Public Health* **8**, 194–201. (doi:10.1016/j.jiph.2014.09.009)
- 49. Frenkel JK, Dubey JP. 1972 Rodents as vectors for feline coccidia, *Isospora felis* and *Isospora rivolta*. J. Infect. Dis. **125**, 69–72. (doi:10.1093/infdis/125. 1.69)
- Guernier V, Hochberg ME, Guégan J-F. 2004 Ecology drives the worldwide distribution of human diseases. *PLoS Biol.* 2, e141. (doi:10.1371/journal. pbio.0020141)
- Cashdan E. 2014 Biogeography of human infectious diseases: a global historical analysis. *PLoS ONE* 9, e0106752. (doi:10.1371/journal.pone.0106752)
- Thieltges DW, Hof C, Dehling DM, Brändle M, Brandl R, Poulin R. 2011 Host diversity and latitude drive trematode diversity patterns in the European freshwater fauna: trematode diversity patterns. *Global Ecol. Biogeogr.* 20, 675–682. (doi:10.1111/j. 1466-8238.2010.00631.x)
- Díaz-Regañón D, Villaescusa A, Ayllón T, Rodríguez-Franco F, Baneth G, Calleja-Bueno L, García-Sancho M, Agulla B, Sainz Á. 2017 Molecular detection of *Hepatozoon* spp. and *Cytauxzoon* sp. in domestic and stray cats from Madrid, Spain. *Parasites Vectors* 10, 112. (doi:10.1186/s13071-017-2056-1)
- Bell ET, Toribio JLML, White JD, Malik R, Norris JM. 2006 Seroprevalence study of feline coronavirus in owned and feral cats in Sydney, Australia. *Aust. Vet. J.* 84, 74–81. (doi:10.1111/j.1751-0813.2006. tb12231.x)
- Chang-Fung-Martel J, Gummow B, Burgess G, Fenton E, Squires R. 2013 A door-to-door prevalence study of feline immunodeficiency virus in an Australian suburb. *J. Feline Med. Surg.* 15, 1070-1078. (doi:10.1177/1098612X13491959)
- Norris JM, Bell ET, Hales L, Toribio J-ALML, White JD, Wigney DI, Baral RM, Malik R. 2007 Prevalence of feline immunodeficiency virus infection in domesticated and feral cats in eastern Australia. *J. Feline Med. Surg.* **9**, 300–308. (doi:10.1016/j. jfms.2007.01.007)
- Ravi M, Wobeser GA, Taylor SM, Jackson ML. 2010 Naturally acquired feline immunodeficiency virus (FIV) infection in cats from western Canada: prevalence, disease associations, and survival analysis. *Can. Vet. J.* **51**, 271–276.
- Walker VR, Morera Galleguillos F, Gómez Jaramillo M, Pereira Almosny NR, Arauna Martínez P, Grob Behne P, Acosta-Jamett G, Müller A. 2016 Prevalence, risk factor analysis, and hematological findings of *Hemoplasma* infection in domestic cats from Valdivia, southern Chile. *Comp. Immunol. Microbiol. Infect. Dis.* 46, 20–26. (doi:10.1016/j. cimid.2016.03.004)
- 59. Bergmann M, Englert T, Stuetzer B, Hawley JR, Lappin MR, Hartmann K. 2017 Risk factors of

royalsocietypublishing.org/journal/rsbl *Biol. Lett.* **15**: 20180840

7

different *Hemoplasma* species infections in cats. *BMC Vet. Res.* **13**, 52. (doi:10.1186/s12917-017-0953-3)

- Baneth G, Sheiner A, Eyal O, Hahn S, Beaufils J-P, Anug Y, Talmi-Frank D. 2013 Redescription of *Hepatozoon felis* (Apicomplexa: Hepatozoidae) based on phylogenetic analysis, tissue and blood form morphology, and possible transplacental transmission. *Parasites Vectors* 6, 102. (doi:10.1186/1756-3305-6-102)
- Meneses IDS *et al.* 2014 Frequency of antibodies against *Sarcocystis neurona* and *Neospora caninum* in domestic cats in the state of Bahia, Brazil. *Rev. Bras. Parasitol. Vet.* 23, 526–529. (doi:10.1590/ S1984-29612014080)
- Nijsse R, Ploeger HW, Wagenaar JA, Mughini-Gras L. 2016 Prevalence and risk factors for patent *Toxocara* infections in cats and cat owners' attitude towards deworming. *Parasitol. Res.* **115**, 4519–4525. (doi:10.1007/s00436-016-5242-8)
- Must K, Lassen B, Jokelainen P. 2015 Seroprevalence of and risk factors for *Toxoplasma gondii* infection in cats in Estonia. *Vector Borne Zoonotic Dis.* 15, 597–601. (doi:10.1089/vbz.2015.1809)
- Ahmad N, Ahmed H, Irum S, Qayyum M. 2014 Seroprevalence of IgG and IgM antibodies and associated risk factors for toxoplasmosis in cats and dogs from sub-tropical arid parts of Pakistan. *Trop. Biomed.* 31, 777–784.
- Deksne G, Petrusēviča A, Kirjušina M. 2013 Seroprevalence and factors associated with *Toxoplasma gondii* infection in domestic cats from urban areas in Latvia. *J. Parasitol.* 99, 48–50. (doi:10.1645/GE-3254.1)
- Györke A, Opsteegh M, Mircean V, Iovu A, Cozma V. 2011 *Toxoplasma gondii* in Romanian household cats: evaluation of serological tests, epidemiology and risk factors. *Prev. Vet. Med.* **102**, 321–328. (doi:10.1016/j.prevetmed.2011.07.015)
- Ketzis JK, Shell L, Chinault S, Pemberton C, Pereira MM. 2015 The prevalence of *Trichuris* spp. infection in indoor and outdoor cats on St. Kitts. *J. Infect. Dev. Ctries* 9, 111–113. (doi:10.3855/jidc.5778)
- Opsteegh M, Haveman R, Swart AN, Mensink-Beerepoot ME, Hofhuis A, Langelaar MFM, van der Giessen JWB. 2012 Seroprevalence and risk factors for *Toxoplasma gondii* infection in domestic cats in The Netherlands. *Prev. Vet. Med.* **104**, 317–326. (doi:10.1016/j.prevetmed.2012.01.003)
- 69. Burling AN, Levy JK, Scott HM, Crandall MM, Tucker SJ, Wood EG, Foster JD. 2017 Seroprevalences of feline leukemia virus and feline immunodeficiency

virus infection in cats in the United States and Canada and risk factors for seropositivity. *J. Am. Vet. Med. Ass.* **251**, 187–194. (doi:10.2460/javma.251. 2.187)

- Higgins JPT, Thompson SG. 2002 Quantifying heterogeneity in a meta-analysis. *Stat. Med.* 21, 1539–1558. (doi:10.1002/sim.1186)
- Day MJ *et al.* 2012 Surveillance of zoonotic infectious disease transmitted by small companion animals. *Emerg. Infect. Dis.* **18**, e1. (doi:10.3201/ eid1812.120664)
- Carver S et al. 2016 Pathogen exposure varies widely among sympatric populations of wild and domestic felids across the United States. *Ecol. Appl.* 26, 367–381. (doi:10.1890/15-0445)
- Higgins JPT, Thompson SG, Spiegelhalter DJ. 2009 A re-evaluation of random-effects meta-analysis. *J. R. Stat. Soc. A* **172**, 137–159. (doi:10.1111/j. 1467-985X.2008.00552.x)
- Johnson PTJ, Hoverman JT. 2012 Parasite diversity and coinfection determine pathogen infection success and host fitness. *Proc. Natl Acad. Sci. USA* 109, 9006–9011. (doi:10.1073/pnas.1201790109)
- Johnson PTJ, Preston DL, Hoverman JT, LaFonte BE. 2013 Host and parasite diversity jointly control disease risk in complex communities. *Proc. Natl Acad. Sci. USA* **110**, 16 916–16 921. (doi:10.1073/ pnas.1310557110)
- Han BA, Kramer AM, Drake JM. 2016 Global patterns of zoonotic disease in mammals. *Trends Parasitol.* 32, 565–577. (doi:10.1016/j.pt.2016.04. 007)
- American Bird Conservancy. Cats indoors. Better for cats, better for birds, better for people. The Plains, VA: American Bird Conservancy. See https://abcbirds. org/program/cats-indoors/ (accessed 4 April 2019).
- People for the Ethical Treatment of Animals. Animal rights uncompromised: 'outdoor cats'. Norfolk, VA: PETA. See https://www.peta.org/issues/animalcompanion-issues/cruel-practices/outdoor-cats/ (accessed 4 April 2019).
- Lepczyk CA, Mertig AG, Liu J. 2004 Landowners and cat predation across rural-to-urban landscapes. *Biol. Conserv.* **115**, 191–201. (doi:10.1016/S0006-3207(03)00107-1)
- Clancy EA, Moore AS, Bertone ER. 2003 Evaluation of cat and owner characteristics and their relationships to outdoor access of owned cats. *J. Am. Vet. Med. Ass.* 222, 1541–1545. (doi:10. 2460/javma.2003.222.1541)

- Lepczyk CA *et al.* 2010 What conservation biologists can do to counter trap-neuter-return: response to Longcore *et al. Conserv. Biol.* 24, 627–629. (doi:10. 1111/j.1523-1739.2009.01426.x)
- Chomel BB. 2014 Emerging and re-emerging zoonoses of dogs and cats. *Animals (Basel)* 4, 434–445. (doi:10.3390/ani4030434)
- Robertson ID, Irwin PJ, Lymbery AJ, Thompson RC. 2000 The role of companion animals in the emergence of parasitic zoonoses. *Int. J. Parasitol.* **30**, 1369–1377. (doi:10.1016/S0020-7519(00)00134-X)
- Hunter PR, Thompson RCA. 2005 The zoonotic transmission of *Giardia* and *Cryptosporidium*. *Int. J. Parasitol.* **35**, 1181–1190. (doi:10.1016/j. ijpara.2005.07.009)
- Moher D, Liberati A, Tetzlaff J, Altman DG, PRISMA Group. 2009 Preferred reporting items for systematic reviews and meta-analyses: the PRISMA statement. *PLoS Med.* 6, e1000097. (doi:10.1371/journal.pmed. 1000097)
- Chalkowski K, Wilson A, Lepczyk C, Zohdy S. 2019 Data from: Who let the cats out: a global metaanalysis on risk of parasitic infection in indoor versus outdoor domestic cats (*Felis catus*). Figshare Digital Repository (https://figshare.com/s/ 3eebaf42e161c0e7e1ef).
- Chalkowski K, Wilson A, Lepczyk C, Zohdy S. 2019 Data from: Who let the cats out: a global metaanalysis on risk of parasitic infection in indoor versus outdoor domestic cats (*Felis catus*). Figshare Digital Repository. See https://figshare.com/s/ a334c7815b128cb63b98.
- Viechtbauer W. 2007 Accounting for heterogeneity via random-effects models and moderator analyses in meta-analysis. *Z.r Psychol. J. Psychol.* 215, 104–121. (doi:10.1027/0044-3409.215.2.104)
- Szumilas M. 2010 Explaining odds ratios. J. Can. Acad. Child Adolesc. Psychiat. 19, 227–229. (doi:10. 1007/s00787-010-0087-7)
- Cuijpers P, van Straten A, Bohlmeijer E, Hollon SD, Andersson G. 2010 The effects of psychotherapy for adult depression are overestimated: a meta-analysis of study quality and effect size. *Psychol. Med.* 40, 211. (doi:10.1017/S0033291709006114)
- Duval S, Tweedie R. 2000 Trim and fill: a simple funnel-plot-based method of testing and adjusting for publication bias in meta-analysis. *Biometrics* 56, 455–463. (doi:10.1111/j.0006-341X. 2000.00455.x)