

REVIEWS

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Effects of cyanobacterial toxicity and morphology on the population growth of freshwater zooplankton: Meta-analyses of laboratory experiments

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Abstract

We synthesized data from 66 published laboratory studies, representing 597 experimental comparisons, examining the effects of cyanobacterial toxicity and morphology on the population growth rate and survivorship of 17 genera (34 species) of freshwater, herbivorous zooplankton. Two meta-analyses were conducted with these data. The primary analysis compared herbivore population growth rates for grazers fed treatment diets containing cyanobacteria versus control diets comprising phytoplankton that are generally considered to be nutritious for zooplankton (chlorophytes and/or flagellates). This analysis confirmed that cyanobacteria were poor foods relative to small chlorophytes and flagellates. More importantly, filamentous cyanobacteria were found to be significantly better foods for grazers than single-celled cyanobacteria over all studies. Surprisingly, the presence or absence of commonly-measured toxic compounds (microcystins in 70% of the cases) in the diet had no overall influence on grazer population growth relative to control diets. A secondary analysis compared survival rates for grazers fed cyanobacteria versus no food. In contrast to the primary analysis, grazer survival was more negatively affected by toxic cyanobacteria than non-toxic cyanobacteria, relative to starvation. However, this difference was attributable to the effects of a single *Microcystis* strain, PCC7820. Thus, though some cyanobacterial strains appear to be toxic to some strains of zooplankton, the overall role of commonly-assayed cyanobacterial toxins as a determinant of food quality may be less than widely assumed. We suggest that more attention be focused on nutritional deficiencies, morphology, and the toxicity of undescribed cyanobacterial compounds as mediators of the poor food quality of cyanobacteria for zooplankton.

Interactions between bloom-forming cyanobacteria and zooplankton are central to the responses of freshwater phytoplankton assemblages to nutrient enrichment and top-down manipulation (Burns 1987; Lampert 1987; Sommer 1989). Consequently, a large body of research

has been directed at understanding mechanisms by which cyanobacteria and zooplankton affect each other. This information has important implications for understanding planktonic community structure and function, as well as water quality, in lakes. For example, the filamentous

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morphology of certain cyanobacteria has been shown to negatively affect large cladocerans more than small cladocerans through reduced fecundity (Webster and Peters 1978; Gilbert 1990), and this mechanism may help to explain decreases in the dominance of large cladocerans with lake eutrophication (Jeppesen et al. 2000). In addition, cyanobacteria produce secondary metabolites that can cause illness and, in some cases, death of livestock and humans exposed to water with high cyanobacterial concentrations (Carmichael et al. 2001). If these metabolites are similarly toxic to copresent zooplankton, the potential for stimulation of toxin production via interactions between cyanobacteria and zooplankton could have implications for public health. Given these considerations, there is considerable value in deciphering the specific mechanisms of interaction between cyanobacteria and zooplankton.

It is well established that bloom-forming cyanobacteria are, in a general sense, of poor food quality for herbivorous zooplankton (Porter and Orcutt 1980). The following three properties of cyanobacteria have been proposed to account for this poor quality: filamentous and/or colonial morphologies, production of intracellular secondary metabolites with toxic properties, and deficiencies in essential nutrients (Porter and Orcutt 1980; Lampert 1987; DeMott 1989). Of these, morphology and toxins have attracted the most interest (Lampert 1982; Fulton and Paerl 1987; Lüring 2003). However, distinguishing between these alternative determinants of food quality is not a simple matter. Many food-quality studies have been limited to simple comparisons of zooplankton performance on diets containing or lacking cyanobacteria (Arnold 1971; Porter and Orcutt 1980), but this design does not provide clear evidence for the role of alternative determinants, given the large variety of physiological and morphological differences between cyanobacteria and other phytoplankters.

Large cyanobacterial forms, such as colonies and filaments, have been shown to mechanically interfere with zooplankton grazing by reducing ingestion of food particles (Webster and Peters 1978; Lynch 1980). An unequivocal test of the role of morphology would involve comparing the performance of zooplankton on diets consisting of cyanobacteria from the same strain but with contrasting morphologies. Such tests have been performed in relatively few studies in which a colonial or filamentous species has been disrupted into single cells or small clusters of cells via shaking or sonication. Some studies show that grazers perform better on diets composed of larger forms of cyanobacteria (Chan et al. 2004), whereas other studies show that smaller forms of cyanobacteria are better food for grazers (Hartmann 1985). These conflicting observations suggest that other factors, such as intracellular toxins and nutrient deficiencies, may interact with cyanobacterial morphology and/or grazer size in affecting zooplankton performance (Lampert 1987).

Unequivocal tests of the role of secondary metabolites are more difficult in practice and consequently rarer still. Cyanobacterial toxins have been hypothesized to be important determinants of the poor quality of cyanobacterial diets, but most studies have relied on additions of dissolved cyanobacterial extracts or purified toxins to

zooplankton media (DeMott et al. 1991; Jungmann 1992), comparisons between a cyanobacterium and a starvation control (Lampert 1981), comparisons between a nutritious alga and a cyanobacterium (Lampert 1981), or comparisons between two strains of the same cyanobacterium species that differ in their ability to produce a well-studied toxin (Smith and Gilbert 1995). In general, these types of comparisons do not provide unequivocal tests of the specific role of cyanobacterial toxins in driving poor zooplankton performance. In the examples specified above, the treatment diets compared may have differed in many ways other than in the presence of a particular toxin, such as nutrient content, size, and morphology. Studies that use a starvation control are limited to comparisons of survival rates and, as a result, do not fully address effects on population growth rate. In addition, tests of the effects of dissolved toxins do not mimic the typical route of exposure for freshwater grazers (via feeding) and have involved toxin exposure levels that are orders of magnitude higher than concentrations typically observed in freshwater lakes (Chorus and Bartram 1999). Recent studies (Rohrlack et al. 1999; Kaebernick et al. 2001; Lüring 2003) comparing cyanobacteria from the same strain that have toxic, wild-type genotypes (which contain several microcystin variants), or nontoxic, mutant genotypes (which are unable to synthesize microcystins) have so far provided less than definitive support for the importance of the most commonly studied class of cyanobacterial toxin, microcystin, as a driver of poor zooplankton performance. For example, Lüring (2003) reported similar population growth rates for grazers that were fed a toxic, wild-type strain of *Microcystis* and grazers that were fed the nontoxic, mutant strain derived from the toxic, wild-type *Microcystis* strain.

In lieu of many direct tests of the effects of cyanobacterial morphology and toxicity on zooplankton performance, we conducted a meta-analysis of the dietary effects of cyanobacteria on zooplankton population growth rates, to quantitatively synthesize the current state of knowledge. In this analysis, we compared population growth rates for zooplankton fed treatment foods containing cyanobacteria with those for zooplankton fed control foods comprising chlorophytes and/or flagellates. We also recognized that some experiments address the issue of cyanobacterial toxicity in a way (e.g., via survivorship studies) that precludes them from being included in this meta-analysis. For these studies, we conducted a supplementary meta-analysis to summarize experiments in which the survival rate for grazers fed toxic or nontoxic cyanobacteria was compared with that for a control group that received no food. Comparison of a group receiving a cyanobacterial diet with a group that is undergoing starvation is a long-standing approach to the assessment of dietary toxicity (Lampert 1981), because it is assumed that grazers will die sooner if they receive toxic food than if they receive no food. Such an experimental design might be open to alternative interpretation if grazers reduce their grazing effort in the absence of food (Plath 1998) and thus have higher survival when starved than when in the presence of nutritionally deficient cyanobacteria.

Given that a wide variety of cyanobacterial taxa with different morphologies and toxicological characteristics have been used in a large number of experiments, meta-analysis provides a powerful means of quantitatively synthesizing this literature (Osenberg et al. 1999) and indirectly assessing the role of these alternative determinants of food quality. We hypothesized that, if morphology and/or toxin production are, in general, important determinants of the poor food quality of cyanobacteria, obvious differences in zooplankton performance should be apparent across these categories in the literature. Too few studies provided detailed information about nutritional composition to permit inclusion of nutritional inadequacy as a determinant of food quality in our analyses. We chose to focus on the population growth rate (r) in assessing zooplankton performance, despite a plethora of performance measures that are cited in the literature, because population growth rate integrates many alternative measures (e.g., feeding, assimilation, somatic growth, survival, and reproduction), determines evolutionary fitness, and provides the most direct link to a major goal of much of this research, which is to understand zooplankton population and community dynamics as they are affected by cyanobacteria.

Methods

We searched the literature for experimental studies of the effect of cyanobacteria on zooplankton performance using Web of Science (from 1945 to 2004), Aquatic Sciences and Fisheries Abstracts (from 1971 to 2004), the reference lists of collected articles, and suggestions by scientists familiar with these types of studies. We included all comparisons that provided data on, or from which could be calculated, population growth rates for freshwater zooplankton fed a control diet (i.e., small chlorophyte and/or flagellated algae) versus rates for those fed a diet containing cyanobacteria (the “treatment” diet) and population growth rates for freshwater zooplankton fed a treatment diet containing toxic or nontoxic cyanobacteria versus rates for those fed no food (the “starvation” diet).

The primary analysis contrasted growth rates for herbivore populations receiving treatment diets with those for herbivore populations receiving control diets, as this was our main interest and many studies used this design. No studies were found that measured population growth rate for copepods fed cyanobacteria-containing foods and a control food; thus, only cladocerans and rotifers were included in the primary analysis. The 43 articles included in the primary meta-analysis comprised 358 experimental comparisons (Web appendix 1, Tables A1 and A3; available at: http://www.aslo.org/lo/toc/vol_51/issue_4/1915a1.pdf) across ≥ 2 dietary treatments, including 19 cladoceran and 8 rotifer species from 8 and 5 genera, respectively (Web appendix 1, Table A3). Twenty-two comparisons from six studies were excluded from the data set (Web appendix 1, Table A4) because treatment r estimates could not be calculated or because control r estimates were near or < 0 , which cannot be accommodated by our metric of effect size.

In most cases (99%), zooplankton population growth rates for grazers fed control foods were reported in

the articles included in the primary analysis. In the few cases in which control r values were not reported (Lundstedt and Brett 1991; Gilbert 1994), we calculated r from initial and final population densities ($r = \frac{\ln density_{t+1} - \ln density_t}{time}$). Most population growth rate estimates (90%) for grazers fed treatment foods also came directly from the studies. However a few studies did not provide estimates of r (Web appendix 1, Fig. A1), presumably because of a lack of reproduction or because all of the experimental animals died during the assays. We did not want to exclude these studies, because they provided worst-case scenarios (i.e., large negative effects of cyanobacteria) for experimental comparisons, so we estimated r for these studies from a lack of reproduction but no mortality ($r = 0$; $n = 4$) and survivorship curves ($n = 33$). For studies in which all grazers died before the end of the experiment, we used a population size of 1 on the day the last grazer died to calculate r .

We included all comparisons regardless of whether estimates of among-replicate variance were provided and conducted an unweighted meta-analysis, to avoid biasing the data set. Use of data-exclusion criteria that are not specifically tied to the data’s relevance to the question or that relate only to study “quality” can lead to spurious conclusions (Englund et al. 1999). However, given that variance-weighted estimates enable more-powerful statistical tests, we also extracted among-replicate variances, when available, for use in a parallel set of variance-weighted meta-analyses. We compared the two sets of analyses to assess the robustness of the results (Englund et al. 1999).

Additional information from each study was recorded to enable categorization of the data by cyanobacterial morphology (single-celled, colonial, or filamentous), cyanobacterial toxicity (nontoxic, toxic, or not determined), zooplankton type (cladoceran or rotifer), adult zooplankton length (actual lengths were provided in most studies; for the remaining studies, estimates were collected from the literature), control food type, carbon concentration of control and treatment foods (carbon concentrations were provided in many studies; for the remaining studies, estimates were determined by calculating biovolume from the cell density and cell dimensions [actual measurements were provided in most studies; for the remaining studies, estimates were collected from the literature] and assuming a specific gravity of 1, a ratio of dry mass to wet mass of 0.10, and a carbon content of dry biomass of 50% (Riemann et al. 1989)), and percentage of cyanobacterial carbon in treatment foods.

The classification of a cyanobacterium as toxic or nontoxic is central to our analysis but is not straightforward. In vitro assays (e.g., ELISA, HPLC, and LC/MS) for chemically described secondary metabolites with an established toxicity toward some organism(s) may miss inhibitory compounds that have yet to be described. Reliance on such assays for classification might result in some toxic cyanobacteria being misclassified as nontoxic. Alternatively, bioassays that use terrestrial animals or aquatic animals with no previous evolutionary history of exposure to the compound may result in a compound being characterized

as toxic when it is not inhibitory to animals that coexist with the compound in nature. In this case, nontoxic strains would be misclassified as toxic. These ambiguities cannot be resolved by any classification scheme. Indeed, one of the main motivations for this work was to assess whether well-known compounds are indeed inhibitory, in a general sense, to zooplankton population growth when they are part of the diet. Consequently, we chose to focus on the following specific question: does the presence or absence of previously described secondary metabolites with toxic effects on some organism have an influence on the population growth rate of zooplankton, in general?

Cyanobacteria were considered toxic if they tested positive via in vitro analysis for known toxins (anatoxin, cylindrospermopsin, deazaadenosine, glucopyranoside, microcystin, microviridin, and paralytic shellfish toxin). Toxin concentrations are known to vary within and among cyanobacterial strains under different environmental conditions and physiological states (Lee et al. 2000). However, these data were not routinely provided and consequently could not be included in our analysis. Nontoxic cyanobacteria were those that either tested negative for known toxins via in vitro analysis or, in one case (Arnold 1971), were considered to be nontoxic because the strains were members of genera (i.e., *Gleocapsa*, *Merismopedia*, and *Synechocystis*) that, to our knowledge, have never been shown to produce toxins. Cases in which potentially toxic genera of cyanobacteria (e.g., *Microcystis*, *Anabaena*, and *Oscillatoria Planktothrix*) were not chemically analyzed for toxins were designated as “not determined.” We contacted authors to obtain information about the toxicity of cyanobacterial strains when this information was not included in their article. This inquiry netted an additional 45 comparisons (total $n=265$) that could be classified as toxic or nontoxic in the primary analysis.

For the primary analysis, the effect of cyanobacteria on zooplankton population growth relative to control foods was calculated for each experimental comparison ($n = 358$) as follows:

$$\text{Unweighted effect size} = \frac{r_t - r_c}{r_c}$$

where r_t is the population growth rate on the cyanobacterial (treatment) diet and r_c is the population growth rate on the control diet (e.g., small chlorophyte and/or flagellate). This metric was chosen to standardize effect sizes by maximum population growth rate (i.e., growth rate on control foods) across taxa with varying growth rates and to center the results about zero to facilitate interpretation. An effect size of zero indicates that growth rates on the two diets were similar. Negative effect sizes indicate that growth rates were lower during receipt of the cyanobacterial diet than growth during receipt of the control diet. Effect sizes less than -1 indicate that population growth during receipt of the cyanobacteria diet (r_t) was negative, because r_c was always greater than zero. Weighted effect sizes ($n = 285$) were calculated as follows:

$$\text{Weighted effect size} = \frac{r_t}{r_c} + \frac{r_t \times \text{var } r_c}{r_c^3} - 1$$

Variances for these weighted effects sizes were calculated as follows:

$$\text{Weighted variance} = \frac{\text{var } r_t}{r_c^2} + \frac{\text{var } r_c}{r_c^4}$$

In the secondary meta-analysis, 23 articles provided data ($n = 239$) for cladocerans (5 genera and 12 species), copepods (3 genera and 3 species), a rotifer (1 genus and 1 species), and a fairy shrimp (1 genus and 1 species) fed nontoxic ($n = 27$) or toxic ($n = 212$) cyanobacteria or no food (Web appendix 1, Tables A2 and A3). Data for one study (Arnold 1971) were not included because starved grazers exhibited positive population growth, suggesting a methodological problem in the starvation treatment (Web appendix 1, Table A4). In this analysis, estimates of r (measured as survival rate) were collected directly or calculated from initial and final densities. For studies in which all grazers died before the end of the experiment, we used a population size of 1 on the day the last grazer died to calculate r .

A different effect size metric was needed for the secondary analysis, because the data in most of these studies were limited to survivorship estimates over time, which result in population growth rate estimates ≤ 0 . Because estimates of $r \leq 0$ cannot be accommodated by the effect size metric noted above ($\frac{r_t - r_c}{r_c}$), we calculated effect sizes for grazers fed cyanobacteria-containing foods (treatment) with effect sizes for starved grazers as follows:

$$\text{Difference effect size day}^{-1} = r_t - r_s$$

where r_t is the survival rate during receipt of the cyanobacterial (treatment) diet, and r_s is the survival rate for starved grazers. An effect size of zero indicates that survival rates associated with the two diets were similar. Negative effect sizes indicate that survival rates during receipt of the cyanobacterial diet were lower than those for the starved diet (i.e., that zooplankton survival was inhibited by cyanobacteria relative to starvation).

Population growth rates for grazers fed control or treatment foods were compared using a paired Student's t -test. Analysis of variance (ANOVA) and its weighted analog (MetaWin, version 2.1) were used to test for differences in effect size among categorical means and to identify significant interactions, and Tukey's multiple comparison tests was used when >2 means were being compared. We used random-effects models for all categorical, weighted analyses because we did not assume a single, true effect size across the disparate studies, which is done for fixed-effects models. Linear regression was used to examine relationships between effect sizes and grazer size or the percentage of cyanobacteria in treatment foods. A general linear model (GLM) was used to test for interactions (i.e., homogeneity of slopes test) between grazer size and cyanobacterial morphology or toxicity. Analysis of covariance (ANCOVA) was used to test for effects of cyanobacterial toxicity and morphology after accounting for variation attributable to continuous variables (i.e., grazer size and percentage of cyanobacteria in the

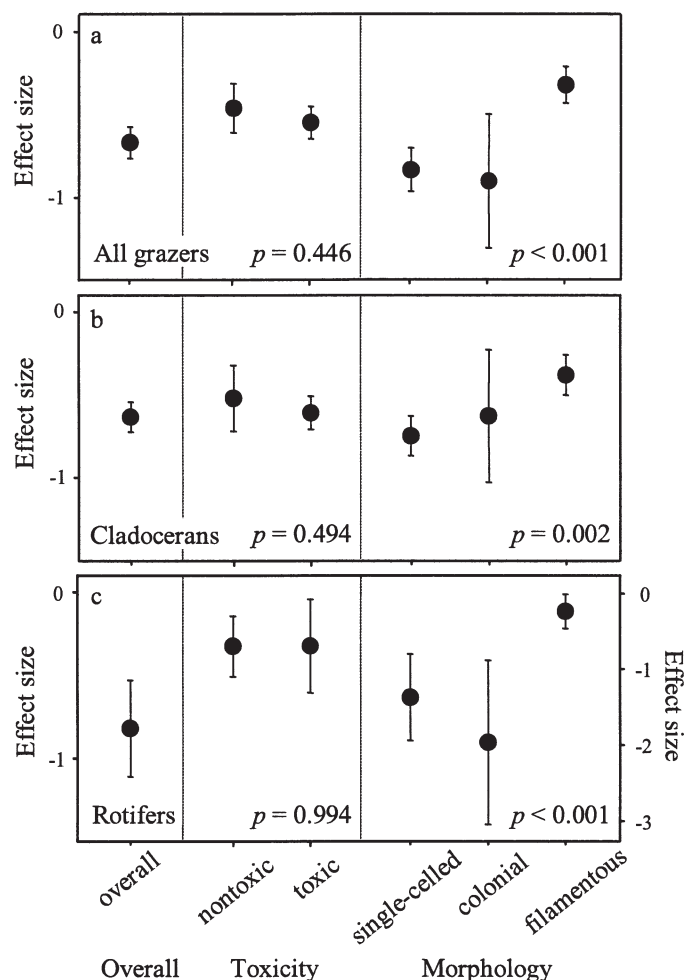


Fig. 1. Effect size averages and 95% confidence intervals for the unweighted data set from the primary meta-analysis comparing r for (a) all grazers, (b) cladocerans only, and (c) rotifers only fed control foods composed of chlorophytes and/or flagellates and treatment foods containing cyanobacteria. Note scale change for cyanobacterial morphology mean effect sizes for rotifers. Sample sizes are presented in Table 1.

treatment diet). ANCOVA was used only if slopes (e.g., effect size vs. grazer length or the percentage of cyanobacteria in the treatment diet) were not significantly different at a p value of <0.05 . Unweighted and weighted analyses were performed with Systat, version 9.01 (SPSS 1998), and MetaWin, version 2.1 (Rosenberg et al. 2002), respectively. The rejection criterion was set a priori at an α level of <0.05 .

Results

As expected, zooplankton population growth rates were much lower ($p < 0.001$, by the paired t -test) for diets containing cyanobacteria (mean [\pm SE], 0.191 ± 0.024), compared with control diets (mean [\pm SE], 0.479 ± 0.025), and as a result, the mean effect sizes averaged over all studies were negative (mean effect sizes were significantly <0 ; $p < 0.001$, by the t test) (Fig. 1; Table 1). The magnitude of

growth inhibition by cyanobacteria also increased as the percentage of cyanobacteria in the treatment diet increased ($p < 0.001$) (Fig. 2a; Table 2). Significant inhibition of zooplankton population growth was evident for all three morphological types (effect sizes for all morphological types were significantly <0 ; $p < 0.001$, by one sample t tests), but single-celled and colonial forms inhibited growth more than did filamentous forms overall (Fig. 1; Table 1). Surprisingly, there was no statistically significant difference in effect size ($p = 0.446$) between populations receiving toxic cyanobacteria and populations receiving nontoxic cyanobacteria, despite large sample sizes (total $n = 265$) and low residual error (Fig. 1; Table 1).

We also examined interactions between grazer length, morphology, and toxicity and cyanobacterial morphology and toxicity. Our analysis of the interaction between morphology and toxicity was limited by the lack of nontoxic colonial species in the data set, and so it only includes a comparison of filamentous and single-celled morphologies. For all grazers, no significant interactions were found between cyanobacterial toxicity ($p = 0.993$) and grazer length ($p = 0.001$) (toxicity \times grazer length interaction, $p = 0.248$) or cyanobacterial morphology ($p < 0.001$) and toxicity ($p = 0.020$) (morphology \times toxicity interaction, $p = 0.313$). However, a significant interaction was found between cyanobacterial morphology ($p < 0.001$) and grazer length ($p = 0.035$) (morphology \times grazer length interaction, $p = 0.004$). Moreover, the effect of cyanobacteria across grazer lengths varied among cyanobacteria types (Table 2). Effect sizes were negatively related to increasing grazer length for filamentous, nontoxic, and toxic cyanobacteria ($p \leq 0.019$) (Table 2) and positively related to increasing grazer length for colonial cyanobacteria ($p = 0.036$) (Table 2), but no relationship was found between grazer length and single-celled cyanobacteria ($p = 0.667$) (Table 2).

There was no statistically significant difference in inhibition between cladocerans and rotifers over all studies ($p = 0.110$) (Fig. 1), which is consistent with the lack of a significant relationship between effect size and grazer length across all studies ($p = 0.428$) (Fig. 2b; Table 2). Moreover, neither type of grazer responded differently to toxic and nontoxic cyanobacteria ($p \geq 0.49$) (Fig. 1; Table 1). However, when analyzed separately, cladocerans and rotifers did respond somewhat differently to the distinct cyanobacterial morphologies ($p \leq 0.002$) (Fig. 1; Table 1). Both grazer groups were inhibited more by single-celled cyanobacteria than by filamentous cyanobacteria, but rotifers were more negatively affected by colonial and single-celled cyanobacteria than were cladocerans ($p \leq 0.005$) (Fig. 1). Of interest, rotifer performance during receipt of filamentous cyanobacteria was similar to their performance during receipt of control foods (Fig. 1; Table 1). However, the mean effect size was negative and nearly significantly <0 (one sample t -test of mean = 0; $p = 0.056$). Unweighted and weighted analyses provided similar outcomes in most of the cases described above (Tables 1 and 2).

Given the unexpected and potentially controversial finding that cyanobacterial toxicity had little influence on

Table 1. Results for the primary meta-analysis comparing r for grazers fed control foods comprised of chlorophytes and/or flagellates and treatment foods containing cyanobacteria.*

Comparisons	Group	Un-weighted n	Unweighted mean effect size	Unweighted 95% CI†	Un-weighted p value‡	Weighted n	Weighted mean effect size	Weighted 95% CI†	Weighted p value‡
Overall effect		358	-0.673	-0.768 to -0.579		285	-0.412	-0.450 to -0.374	
Toxicity	nontoxic	45	-0.457	-0.606 to -0.308	0.446	34	-0.368	-0.463 to -0.272	0.355
	toxic	220	-0.544	-0.642 to -0.447		193	-0.320	-0.360 to -0.281	
Morphology	single-celled	199	-0.836 ^a	-0.967 to -0.704	0.000	155	-0.612	-0.669 to -0.556	0.001
	colonial	38	-0.905 ^a	-1.309 to -0.501		28	-0.568	-0.735 to -0.400	
	filamentous	119	-0.325 ^b	-0.436 to -0.215		101	-0.252	-0.298 to -0.207	
Grazer type	cladocerans	280	-0.633	-0.723 to -0.542	0.110	220	-0.386	-0.432 to -0.340	0.038
	rotifers	78	-0.819	-1.109 to -0.529		65	-0.473	-0.542 to -0.403	
Cladocerans only									
Toxicity	nontoxic	32	-0.513	-0.712 to -0.315	0.494	23	-0.401	-0.517 to -0.285	0.280
	toxic	177	-0.600	-0.700 to -0.500		152	-0.335	-0.382 to -0.288	
Morphology	single-celled	170	-0.747 ^a	-0.866 to -0.629	0.002	130	-0.599	-0.658 to -0.541	0.000
	colonial	30	-0.627 ^{a,b}	-1.026 to -0.228		24	-0.372	-0.560 to -0.184	
	filamentous	78	-0.380 ^b	-0.502 to -0.258		66	-0.195	-0.241 to -0.150	
Rotifers only									
Toxicity	nontoxic	13	-0.318	-0.500 to -0.136	0.994	11	-0.292	-0.490 to -0.093	0.951
	toxic	43	-0.316	-0.598 to -0.033		41	-0.286	-0.366 to -0.205	
Morphology	single-celled	29	-1.354 ^a	-1.921 to -0.786	0.000	25	-0.683	-0.845 to -0.522	0.000
	colonial	8	-1.947 ^a	-3.026 to -0.868		4	-1.420	-2.086 to -0.755	
	filamentous	41	-0.221 ^b	-0.447 to 0.006		36	-0.336	-0.459 to -0.213	

* Differences in letter (a,b) indicate significant differences as detected by Tukey's multiple comparison tests.

† Confidence intervals (CIs) containing zero signify that the treatment and control foods affected grazers similarly.

‡ p values relate to specific effect size comparisons (e.g., cladocerans versus rotifers, nontoxic versus toxic cyanobacteria).

mean effect size in studies comparing a cyanobacterial diet with a control diet, we analyzed the primary data set more intensively to examine the robustness of this specific result. First, we examined whether there was an interaction between the concentration of cyanobacteria in treatment foods and the presence or absence of toxicity. There was no interaction between cyanobacterial concentration and toxicity ($p = 0.477$) and no statistically significant effect of toxicity after accounting for variation in cyanobacterial concentration (for percentage of cyanobacteria in the diet, $p < 0.001$, by ANCOVA; for toxicity, $p = 0.378$, by ANCOVA). Second, we restricted the data set to studies in which the treatment diet was made up of 100% cyanobacteria, given that these treatments showed the strongest negative effects on population growth (Fig. 2a). Again, effect sizes between toxic and nontoxic cyanobacteria diets were not significantly different for all grazers ($p = 0.354$), nor for cladocerans ($p = 0.527$) and rotifers ($p = 0.355$) analyzed separately. Finally, we examined the influence of variation in the control diet on effect size. Zooplankton exhibited greater population growth on control diets made up of *Cryptomonas*, *Nannochloris*, or *Euglena* organisms relative to other species (overall, $p < 0.001$) (Fig. 2a; Web appendix 1). We analyzed the data after subdividing effect sizes into two categories on the basis of the type of control food that was used (*Cryptomonas*, *Euglena*, and *Nannochloris* organisms were considered to be better food, and *Chlorella*, *Scenedesmus* and other genera were considered to be poorer food). Neither data subset revealed statistically significant differences in mean effect size for grazers fed toxic or nontoxic cyanobacteria ($p \geq 0.473$).

In direct contrast to the lack of influence of toxicity in the analysis of experiments in which control foods were used (Fig. 1; Table 1), the secondary analysis of experiments that used a starvation treatment revealed that receipt of toxic cyanobacteria (mean [\pm SE], $-0.237 \text{ day}^{-1} \pm 0.027 \text{ day}^{-1}$; $n = 212$) resulted in greater zooplankton growth than did receipt of nontoxic cyanobacteria (mean [\pm SE], $0.093 \text{ day}^{-1} \pm 0.030 \text{ day}^{-1}$; $n = 27$; $p < 0.001$) (Fig. 3). Grazers fed toxic cyanobacteria exhibited significantly lower survival rates than starved animals (one sample t -test of mean = 0; $p < 0.001$), whereas grazers fed nontoxic cyanobacteria exhibited significantly higher survival rates than starved animals (one sample t -test of mean = 0; $p = 0.005$). This data set was heavily biased towards the use of *Microcystis* species (93% of the cyanobacterial treatment food types in the secondary analysis), and more specifically, the microcystin-containing strain *M. aeruginosa* PCC7820 (37% of the toxic treatment foods in the secondary meta-analysis, compared with only 21% of the toxic treatment foods in the larger, primary analysis). Repeated use of the same species or genotype is an obvious source of nonindependence in meta-analysis (Gurevitch and Hedges 1999) and may limit the domain of relevance of our synthesis in much the same way that pseudoreplication affects the interpretation of an individual experiment (Hurlbert 1984). It is possible that the significant influence of toxicity in the secondary analysis is driven solely by the effect of this single genotype of *M. aeruginosa*. To address this possibility, we examined the influence of toxicity in experiments that used PCC7820 and experiments that used other toxic strains (Fig. 3). This analysis revealed that the influence of toxicity on zooplank-

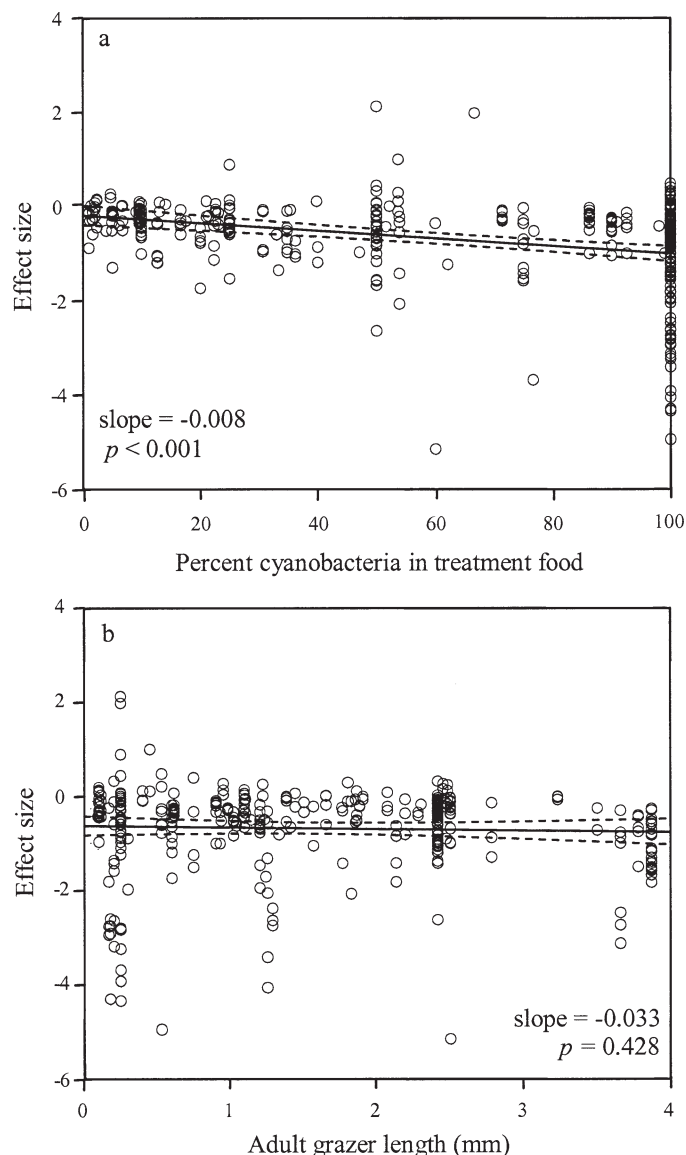


Fig. 2. Relationships between unweighted effect size and (a) percentage of cyanobacteria in treatment foods or (b) adult grazer length (mm) for the primary data set comparing r for grazers fed control foods comprised of chlorophytes and/or flagellates and treatment foods containing cyanobacteria. Solid lines are best-fit lines. Dashed lines are 95% confidence bands. Transformation to remove heteroscedasticity did not change the results of the regression appreciably.

ton survival rates is statistically significant, even in the absence of PCC7820 (results for nontoxic diets are presented above; the mean value [\pm SE] for all other toxic strains besides PCC7820 [$n = 133$] was $-0.184 \text{ day}^{-1} \pm 0.031 \text{ day}^{-1}$ [$p < 0.001$] for all other toxic strains vs. nontoxic strains; the mean value for PCC7820 strains only [$n = 79$] was $-0.326 \text{ day}^{-1} \pm 0.051 \text{ day}^{-1}$ [$p < 0.001$] for PCC7820 strains vs. nontoxic strains), and also that the magnitude of the difference in effect sizes between comparisons of toxic and nontoxic cyanobacteria is much (44%) less for all other strains than it is for PCC7820 (Fig. 3).

Discussion

Using two meta-analyses of published data, we confirmed that cyanobacteria are generally poor food for freshwater grazers, compared with small chlorophytes and/or flagellates, a generalization that is widely presumed to be true (Porter and Orcutt 1980; Sommer 1989; Gilbert 1990). The very low p values associated with the general effect of cyanobacteria and for the effect of increasing concentrations of cyanobacteria ($p < 0.001$) indicate that our analysis was relatively powerful, as would be expected, given the generally large sample sizes (Tables 1 and 2; Web appendix 1, Tables A1, A2, and A3). Despite these large sample sizes, however, we did not find an overall statistical difference in population growth inhibition between toxic and nontoxic cyanobacteria (as defined in our analysis), a generalization that is also widely presumed to be true. We also found that cladocerans and rotifers were affected differently by the distinct cyanobacterial morphologies, a finding that has not been widely recognized before (Gilbert 1990; Nandini and Rao 1998).

In general, cladocerans and rotifers responded similarly to cyanobacteria regardless of toxin absence or presence, compared with control foods. These findings were somewhat unexpected, given that cladocerans and rotifers use different chemosensory and mechanosensory structures to locate and ingest prey (Larsson and Dodson 1993; Snell 1998). However, we did find statistically significant differences between the two grazer groups in their responses to different cyanobacterial morphologies (Fig. 1). Cladocerans and rotifers both grew best on filamentous forms, but rotifers were more inhibited by single-celled and colonial cyanobacteria, whereas cladocerans were most inhibited when fed single-celled cyanobacteria ($p < 0.05$, by Tukey's test) (Fig. 1; Table 1).

Across 27 species of freshwater grazers incorporated in the primary analysis (Web appendix 1, Table A3), filamentous algae were a better food source than single-celled cyanobacteria, compared with control foods. This finding is surprising, given that filamentous cyanobacteria have been shown to mechanically interfere with herbivore-filtering appendages (Lynch 1980; Porter and McDonough 1984; Gliwicz 1990), thus reducing ingestion of food particles. We consider two explanations for the difference between the effects of filamentous and single-celled cyanobacteria. First, for phylogenetic reasons, filamentous and single-celled genera may lack different essential nutrients. For example, the filamentous cyanobacterium *Anabaena* has been found to be deficient in sterols (von Elert et al. 2003), whereas a strain of single-celled *Synechococcus* lacked essential fatty acids (von Elert and Wolffrom 2001). Alternatively, single-celled cyanobacteria may be ingested more readily than filamentous algae, because of grazer gape limitations (Gliwicz and Siedlar 1980), and these smaller prey may contain undescribed secondary metabolites that function as toxins only when consumed (Wolfe 2000).

Single-celled cyanobacteria occur in nature (e.g., *Chroococcus*, *Dactylococcopsis*, *Synechococcus*, and *Synechocystis*), but we are unaware of any naturally-occurring, toxic,

Table 2. Linear regression output for the unweighted and weighted analyses of the primary data set comparing r for grazers fed control foods comprised of chlorophytes and/or flagellates and treatment foods containing cyanobacteria.

Independent variable	Dependent variable	Unweighted n	Unweighted slopes	Unweighted p value	Weighted n	Weighted slope	Weighted p value
Percent cyanobacterial carbon in treatment diet (%)	all grazers	358	-0.008	0.000	285	-0.005	0.000
	cladocerans	280	-0.005	0.000	220	-0.003	0.000
	rotifers	78	-0.020	0.000	65	-0.013	0.000
	nontoxic	45	-0.008	0.000	34	-0.011	0.000
	toxic	220	-0.006	0.000	193	-0.002	0.000
	single-celled	199	-0.010	0.000	155	-0.012	0.000
	colonial	38	-0.017	0.022	28	-0.012	0.018
	filamentous	119	-0.005	0.000	102	-0.001	0.034
Grazer adult size (mm)	all grazers	355	-0.033	0.428	282	-0.036	0.035
	nontoxic	45	-0.151	0.003	34	-0.139	0.007
	toxic	217	-0.243	0.000	190	-0.092	0.000
	single-celled	199	0.024	0.667	155	-0.061	0.102
	colonial	38	0.605	0.036	28	0.412	0.060
	filamentous	116	-0.142	0.019	99	0.057	0.002

single-celled, freshwater cyanobacterial species. Given that species of *Microcystis*, a strictly colonial genus (Komárek 1991), constituted 62% of the treatment foods types in our primary meta-analysis, our observation that relatively few studies measured population growth for grazers fed colonial cyanobacteria is surprising. However, as near as we can ascertain, *Microcystis* strains from culture collections almost invariably grow as single cells, and many (35%) of the articles in our primary meta-analysis used culture-collection strains from this genus. A total of 83% of the effect size comparisons involving *Microcystis* species used single-celled strains. Given the importance of morphology as a determinant of food quality (Fig. 1; Tables 1

and 2) and grazing-resistance (Sterner 1989), widespread use of single-celled strains of colonial species in studies of phytoplankton-zooplankton interactions is problematic. This problem can be overcome by using cyanobacteria that have been recently collected from the field (Nandini and Rao 1998) or remain colonial in culture (Ferrão-Filho and Azevedo 2003), whereas *Microcystis* can retain the colonial habit for several years (A.E. Wilson, pers. comm.).

We did not find any evidence that chemically assayed secondary metabolites, such as anatoxins and microcystins, had an overall influence on population growth rates of cladocerans and rotifers (Fig. 1), but we did find some evidence of a toxicity influence on zooplankton survival rates in the secondary analysis of experiments comparing cyanobacterial diets with starvation diets (Fig. 3). The magnitude of the latter effect, however, was greatly influenced by a single genotype of *M. aeruginosa* (PCC7820), which has been used repeatedly in published studies and has been shown to produce at least 10 different microcystin variants, including microcystin-LR, -LY, -LW, and -LF (Robillot et al. 2000). In fact, if we reduce nonindependence in the secondary data set by representing each genotype by a single observation (by averaging effects sizes within each genotype), the influence of toxicity is no longer statistically significant (the mean value [\pm SE] for nontoxic diets [$n = 7$] was $0.033 \text{ day}^{-1} \pm 0.075 \text{ day}^{-1}$, and the mean value [\pm SE] for toxic diets [$n = 21$] was $-0.209 \text{ day}^{-1} \pm 0.080 \text{ day}^{-1}$; $p = 0.109$). As a consequence, we are forced to conclude that, with perhaps the exception of *M. aeruginosa* genotype PCC7820, there is no strong evidence in the literature to support the generalization that the presence or absence of described cyanotoxins is an important factor driving the poor quality of cyanobacteria as food for zooplankton in general. This does not negate specific cases where zooplankton have been shown to be more negatively affected by diets containing toxic cyanobacteria than by diets containing nontoxic cyanobacteria (Rohrlack et al. 2001), but it suggests that such cases are the exception rather than the rule.

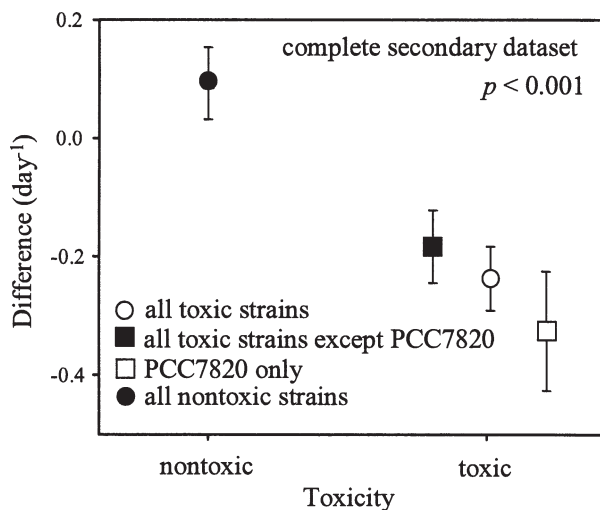


Fig. 3. Effect size differences (day^{-1} , metric: $r_t - r_s$ where r_t is the survival rate on the cyanobacterial (treatment) diet and r_s is the survival rate with no food) and 95% confidence intervals for the secondary analysis. Toxic cyanobacterial data presented as all toxic strains, all toxic cyanobacteria excluding PCC7820, and only data for PCC7820. Complete secondary analysis refers to all nontoxic strains versus all toxic strains.

The potential discrepancy in the influence of cyanobacterial toxicity between the population-growth analyses (Fig. 1) and survival-only analyses (Fig. 3) might be explained by the extraordinary negative effects of cyanobacteria used in the studies included in the secondary analysis (Web appendix 1, Table A2). In these studies, zooplankton fed diets containing toxic cyanobacteria perished before reproduction, which precluded the ability to calculate population growth rate. Studies incorporated in the primary analyses may have used less toxic strains of cyanobacteria or toxic strains at lower doses, thereby enabling the grazers to reproduce. Thus, the inconsistency between the primary and secondary analyses could be attributable to the effects of a small number of strains of toxic cyanobacteria (e.g., PCC7820) and so may not be a general phenomenon (Fig. 3).

Several studies suggest that alternative, undescribed secondary metabolites may be responsible for chemically mediated cyanobacterial inhibition of grazers (Jungmann 1992; Rohrlack et al. 1999). Given the results we report (Fig. 1; Tables 1 and 2), perhaps more research effort should be used to investigate these alternative compounds. In addition, new experimental techniques are needed to better clarify the role of both known and unknown chemical defenses in phytoplankton-zooplankton interactions (Caldwell et al. 2004). As one example, we caution against relying on mouse bioassays (Carmichael 1992) to classify cyanobacterial taxa or secondary metabolites as “toxic” in the ecological sense of a chemical defense against herbivores, because effects on naive vertebrates (e.g., mice) may have little relevance with respect to effects on naturally occurring invertebrates (e.g., zooplankton). Such bioassays are, of course, appropriate for identifying toxins in the context of public health (Carmichael et al. 2001). Finally, recent studies indicate that grazers can adapt, in an evolutionary sense, to the presence of microcystin-containing cyanobacteria in the diet (Hairston 1999; Sarnelle and Wilson 2005). Thus, it also seems prudent to be more circumspect about the source of the zooplankton strains used in laboratory studies of cyanobacteria-zooplankton interactions, if the goal is to understand these interactions in nature. Few of the studies in our meta-analysis provided detailed information about the habitat from which the zooplankton strains investigated were isolated. Given that prolonged exposure to the selective pressures of the laboratory-culture environment may lead to population evolution (Lynch et al. 1991), serious attention should also be paid to the interval between isolation from nature and use in experiments for both phytoplankton and zooplankton strains.

Cyanobacteria-zooplankton interactions can significantly influence planktonic community dynamics following nutrient and fish manipulations (Sommer 1989). With this in mind, understanding the mechanisms mediating these interactions is critical to effective water quality management. Furthermore, our quantitative survey of the cyanobacteria-zooplankton literature highlights a further need for experiments aimed at teasing apart the effects of cyanobacterial morphology, toxicity, nutrient deficiencies, and interactions among these mechanisms on zooplankton

fitness. Finally, copepods are an important link in aquatic food webs (Sommer 1989), and numerous studies have demonstrated the effects of blue-green algae on copepod filtration rate, egg production, and survival (DeMott et al. 1991). Surprisingly, we found only one article that compared individual fitness data for copepods fed a mixture of a cyanobacterium of unknown toxicity and a chlorophyte with fitness data for copepods fed a unialgal diet of the green alga (Twombly et al. 1998). In this study, the cyanobacterial diet was not shown to affect individual grazer fitness. Thus, we encourage future studies aimed at evaluating the effect of cyanobacteria on copepod fitness.

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Effects of cyanobacterial toxicity and morphology on the population growth of freshwater zooplankton: Meta-analyses of laboratory experiments

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Web Appendix 1

Table A1. List of studies used in the primary meta-analysis comparing r for grazers fed control foods comprising chlorophytes and/or flagellates and treatment foods containing cyanobacteria. Grazer types: cl = cladoceran (genera = ¹*Bosmina*, ²*Ceriodaphnia*, ³*Chydoris*, ⁴*Daphnia*, ⁵*Moina*, ⁶*Moinadaphnia*, ⁷*Scapholeberis*, ⁸*Simocephalus*); and r = rotifer (genera = ⁹*Asplanchna*, ¹⁰*Brachionus*, ¹¹*Hexarthra*, ¹²*Keratella*, ¹³*Synchaeta*). Toxins analyzed and found to be present: ¹⁴anatoxin, ¹⁵deazaadenosine, ¹⁶glucopyranoside, ¹⁷microcystin. Morphology types: colonial = colonial [genera = ¹⁸*Microcystis*], filament = filamentous [genera = ¹⁹*Anabaena*, ²⁰*Aphanizomenon*, ²¹*Cylindrospermopsis*, ²²*Oscillatorial Planktothrix*], and single = single-celled (genera = ¹⁹*Anabaena*, ²³*Anacystis*, ²⁴*Gleocapsa*, ²⁵*Merismopedia*, ¹⁸*Microcystis*, ²⁶*Synechococcus*, ²⁷*Synechocystis*). ND = toxin presence not determined by chemical analysis. * Studies employing toxic *Microcystis* PCC7820.

Studies	Grazer type	Cyanobacterial toxicity	Cyanobacterial morphology	Unweighted estimate	Weighted estimate
Ahlgren et al. 1990	cl ^{3,4}	ND	single ¹⁸ , filamentous ²²	4	0
Alva-Martínez et al. 2001	cl ^{2,4,5}	ND	single ¹⁸	9	9
Alva-Martínez et al. 2004	cl ⁴	ND	single ¹⁸	8	8
Arnold 1971	cl ⁴	nontoxic, ND	single ^{19,23,24,25,26,27}	14	0
Brett 1993	cl ⁴	ND	single ¹⁸	2	2
Cecchine 1997	r ¹⁰	toxic ¹⁷	single ¹⁸	9	9
Chen and Xie 2003	cl ⁵	ND	colonial ¹⁸	3	0
Chen and Xie 2004*	cl ⁴	toxic ¹⁷	single ¹⁸	4	0
Claska and Gilbert 1998	cl ⁴	toxic ^{14,15,16}	filament ¹⁹	25	23
de Bernardi et al. 1981	cl ⁴	ND	single ¹⁸	6	0
Ferrão-Filho and Azevedo 2003	cl ^{2,5}	toxic ¹⁷	single ¹⁸ , colonial ¹⁸	16	16
Ferrão-Filho et al. 2000	cl ^{2,4,5,6}	toxic ¹⁷	single ¹⁸	25	23
Ferrão-Filho et al. 2002	cl ^{2,5}	toxic ¹⁷	single ¹⁸	2	2
Fulton and Paerl 1987	r ¹⁰	ND	single ¹⁸ , colonial ¹⁸	4	0
Fulton and Paerl 1988	cl ⁴ , r ¹⁰	ND	colonial ¹⁸	4	0
Gilbert 1990	cl ^{1,2,4} , r ^{12,13}	toxic ^{15,16}	filament ¹⁹	22	22
Gilbert 1994	r ¹²	toxic ¹⁴	filament ¹⁹	1	0
Gilbert 1996a	r ¹⁰	toxic ¹⁴	filament ¹⁹	5	4
Gilbert 1996b	r ^{9,10}	toxic ¹⁴	filament ¹⁹	6	6
Gilbert and Durand 1990	cl ⁴ , r ¹²	nontoxic	filament ¹⁹	4	4
Hanazato and Yasuno 1987	cl ⁵	ND	single ¹⁸ , colonial ¹⁸	2	0
Henning et al. 1991	cl ⁴	toxic ¹⁷ , nontoxic, ND	single ¹⁸	4	0
Hietala et al. 1995*	cl ⁴	toxic ¹⁷	single ¹⁸	10	8
Hietala et al. 1997a*	cl ⁴	toxic ¹⁷	single ¹⁸	18	15
Hietala et al. 1997b*	cl ⁴	toxic ¹⁷	single ¹⁸	4	4
Kurmayer 2001	cl ^{1,4}	nontoxic	filament ²⁰	4	4
Lundstedt and Brett 1991	cl ⁴	ND	single ¹⁸	1	0
Lürling 2003a	cl ⁴	toxic ¹⁷ , nontoxic	single ¹⁸	8	8
Lürling 2003b*	cl ⁴	toxic ¹⁷ , nontoxic	single ¹⁸	16	16
Lürling and van der Grinten 2003	cl ⁴	toxic ¹⁷ , nontoxic	single ¹⁸	5	5
Nandini and Rao 1998	cl ^{2,4,5,7,8} , r ^{10,11}	ND	single ¹⁸ , colonial ¹⁸	32	32
Nandini et al. 2000	cl ⁴	ND	colonial ¹⁸	2	2
Porter and Orcutt 1980	cl ⁴	toxic ¹⁴	single ¹⁹ , filament ¹⁹	4	0
Repka 1996	cl ⁴	toxic ¹⁷	filament ²²	6	0
Repka 1997	cl ⁴	toxic ¹⁷	filament ²²	9	9
Repka 1998	cl ⁴	toxic ¹⁷	filament ²²	12	12
Rothhaupt 1991	r ¹⁰	ND	single ¹⁸ , filament ^{19,21}	8	5
Sartonov 1995	cl ⁴ , r ¹²	nontoxic	single ¹⁸	4	4
Shurin and Dodson 1997	cl ⁴	toxic ¹⁷	single ¹⁸	3	3
Smith and Gilbert 1995*	cl ⁴ , r ¹²	toxic ¹⁷ , nontoxic	single ¹⁸	18	17
Starkweather 1981	r ¹⁰	nontoxic	filament ¹⁹	2	0
Starkweather and Kellar 1983	r ¹⁰	toxic ¹⁴	filament ¹⁹	3	3
Weithoff and Walz 1995	r ¹⁰	toxic ¹⁷	filament ²²	10	10
			Total	358	285

Table A2. List of studies used in the secondary meta-analysis comparing r derived from survivorship curves, LT_{50} estimates, or directly from papers for grazers fed treatment foods containing cyanobacteria and no food. Data types: survivor = survivorship curves, LT_{50} = LT_{50} estimates, and $r = r$ provided in or calculated from paper. Grazer types: ana = anostracan [Genus = ¹*Thamnocephalus*], cl = cladoceran [Genera = ²*Bosmina*, ³*Ceriodaphnia*, ⁴*Daphnia*, ⁵*Moina*, ⁶*Moinadaphnia*], cop = copepod [Genera = ⁷*Cyclops*, ⁸*Diaptomus*, ⁹*Eudiaptomus*], and r = rotifer [Genus = ¹⁰*Brachionus*]. Toxins analyzed and found to be present: ¹¹anatoxin, ¹²cylindrospermopsin, ¹³deazaadenosine, ¹⁴glucopyranoside, ¹⁵microcystin, ¹⁶microviridin, ¹⁷paralytic shellfish toxin]. Morphology types: filament = filamentous [Genera = ¹⁸*Anabaena*, ¹⁹*Aphanizomenon*, ²⁰*Cylindrospermopsis*], and single = single-celled [Genera = ¹⁸*Anabaena*, ²¹*Microcystis*, ²²*Planktothrix*]. ND = toxin presence not-determined by chemical analysis. * = studies employing toxic *Microcystis* PCC7820.

Studies	Data type	Grazer types	Cyanobacterial toxicity	Cyanobacterial morphology	Treatment food – no food
Cecchine 1997	r	r ¹⁰	toxic ¹⁵	single ²¹	1
DeMott et al. 1991*	survivor	cl ⁴ , cop ⁸	toxic ¹⁵ , nontoxic	single ²¹	12
Ferrão-Filho et al. 2000*	survivor	cl ^{3,4,5,6}	toxic ¹⁵	single ²¹	97
Fulton 1988*	survivor	cl ^{2,4,5}	toxic ^{11,15}	single ²¹ , filament ¹⁸	8
Gilbert 1998	survivor	cl ⁴	toxic ^{13,14}	filament ¹⁸	2
Henning et al. 1991	r	cl ⁴	toxic ¹⁵ , nontoxic	single ²¹	2
Henning et al. 2001	survivor	cl ⁴	toxic ¹⁵ , nontoxic	single ²¹	5
Kaebnick et al. 2001	survivor	cl ⁴	toxic ¹⁵ , nontoxic	single ²¹	4
Kurmayer and Jüttner 1999	survivor	ana ¹ , cl ⁴ , cop ^{7,9}	toxic ¹⁵	single ²²	4
Lürling 2003a	r	cl ⁴	toxic ¹⁵ , nontoxic	single ²¹	8
Matveev et al. 1994	survivor	cl ⁴	toxic ¹⁵	single ²¹	1
Nizan et al. 1986*	survivor	cl ⁴	toxic ¹⁵	single ²¹	4
Nogueira et al. 2004a	survivor	cl ⁴	toxic ¹⁷	filament ¹⁹	1
Nogueira et al. 2004b	survivor	cl ⁴	toxic ¹² , nontoxic	filament ²⁰	2
Porter and Orcutt 1980	r	cl ⁴	toxic ¹¹	single ¹⁸ , filament ¹⁸	4
Reinikainen et al. 1994*	survivor	cl ⁴	toxic ¹⁵	single ²¹	48
Rohrlack et al. 1999a	survivor	cl ⁴	toxic ¹⁵ , nontoxic	single ²¹	2
Rohrlack et al. 1999b	LT_{50}	cl ⁴	toxic ¹⁵ , nontoxic	single ²¹	6
Rohrlack et al. 2001	LT_{50}	cl ⁴	toxic ¹⁵ , nontoxic	single ²¹	12
Rohrlack et al. 2004	survivor	cl ⁴	toxic ¹⁶	single ²¹	2
Smith and Gilbert 1995	r	cl ⁴	nontoxic	single ²¹	1
Thostrup and Christoffersen 1999	survivor	cl ⁴	toxic ¹⁵	single ²¹	1
van der Grinten et al. 2000	survivor	cl ⁴	toxic ¹⁵ , nontoxic	single ²¹	12
				Total	239

Table A3. Grazer genera included in the primary and secondary meta-analyses comparing r for grazers fed control foods comprised of chlorophytes and/or flagellates and treatment diets containing cyanobacteria.

Meta-analysis	Genera	Grazer group	No. species	Unweighted effect size estimates
Primary	<i>Asplanchna</i>	rotifer	1	3
Primary	<i>Bosmina</i>	cladoceran	1	3
Primary	<i>Brachionus</i>	rotifer	2	50
Primary	<i>Ceriodaphnia</i>	cladoceran	2	34
Primary	<i>Chydoris</i>	cladoceran	1	2
Primary	<i>Daphnia</i>	cladoceran	10	200
Primary	<i>Hexarthra</i>	rotifer	1	4
Primary	<i>Keratella</i>	rotifer	3	19
Primary	<i>Moina</i>	cladoceran	2	27
Primary	<i>Moinadaphnia</i>	cladoceran	1	6
Primary	<i>Scapholeberis</i>	cladoceran	1	4
Primary	<i>Simocephalus</i>	cladoceran	1	4
Primary	<i>Synchaeta</i>	rotifer	1	2
Primary		Total	27	358
Secondary	<i>Bosmina</i>	cladoceran	1	2
Secondary	<i>Brachionus</i>	rotifer	1	1
Secondary	<i>Ceriodaphnia</i>	cladoceran	1	16
Secondary	<i>Cyclops</i>	copepod	1	1
Secondary	<i>Daphnia</i>	cladoceran	8	164
Secondary	<i>Diaptomus</i>	copepod	1	4
Secondary	<i>Eudiaptomus</i>	copepod	1	1
Secondary	<i>Moina</i>	cladoceran	1	21
Secondary	<i>Moinadaphnia</i>	cladoceran	1	28
Secondary	<i>Thamnocephalus</i>	anostracan	1	1
Secondary		Total	17	239

Table A4. List of excluded comparisons not used in the primary and secondary meta-analyses.

Meta-analysis	Study	Comparisons excluded	Reason for exclusion
Primary	Ahlgren et al. 1990	2	control r near or below 0
Primary	Chen and Xie 2003	9	control r near or below 0
Primary	Ferrão-Filho et al. 2000	4	unable to calculate treatment r
Primary	Laurén-Määttä et al. 1997	4	control r near or below 0
Primary	Lundstedt and Brett 1991	2	control r near or below 0
Primary	Weithoff and Walz 1995	1	control r near or below 0
Primary	Total	22	
Secondary	Arnold 1971	14	starvation r above 0
Secondary	Total	14	

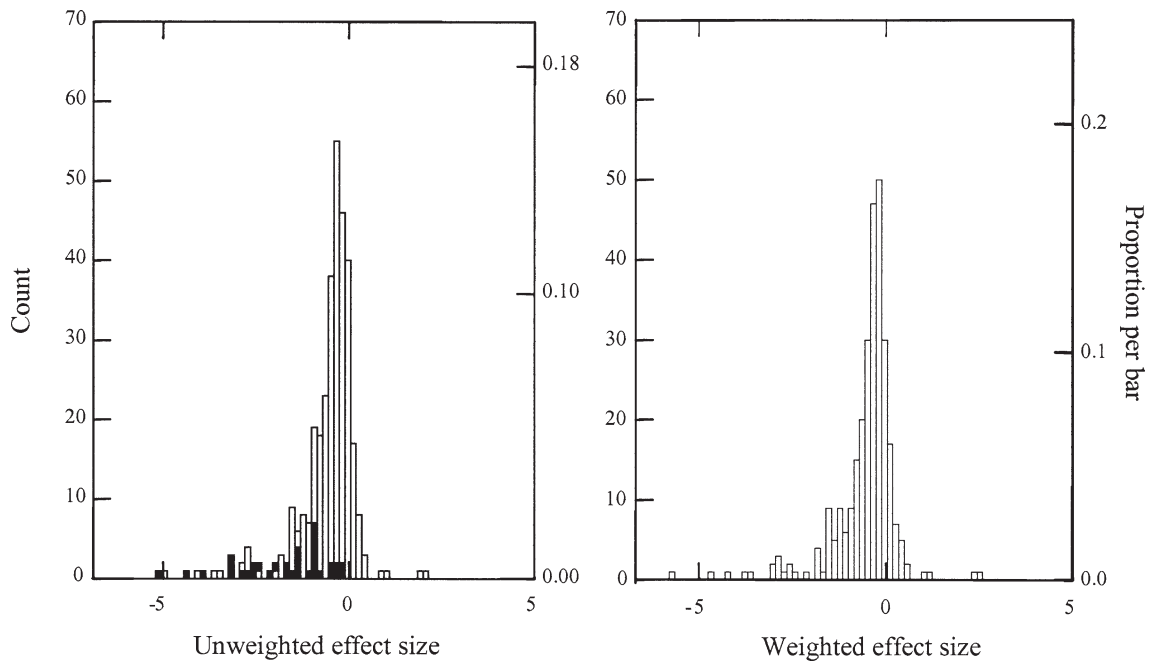


Figure A1. Frequency distribution of effect sizes for unweighted and weighted estimates of r comparisons for grazers fed control foods composed of chlorophytes and/or flagellates and treatment foods containing cyanobacteria used in the primary meta-analysis. Most effect sizes were calculated by using r estimates collected directly from the studies (white bars); however, a few r estimates were calculated from survivorship data (black bars).

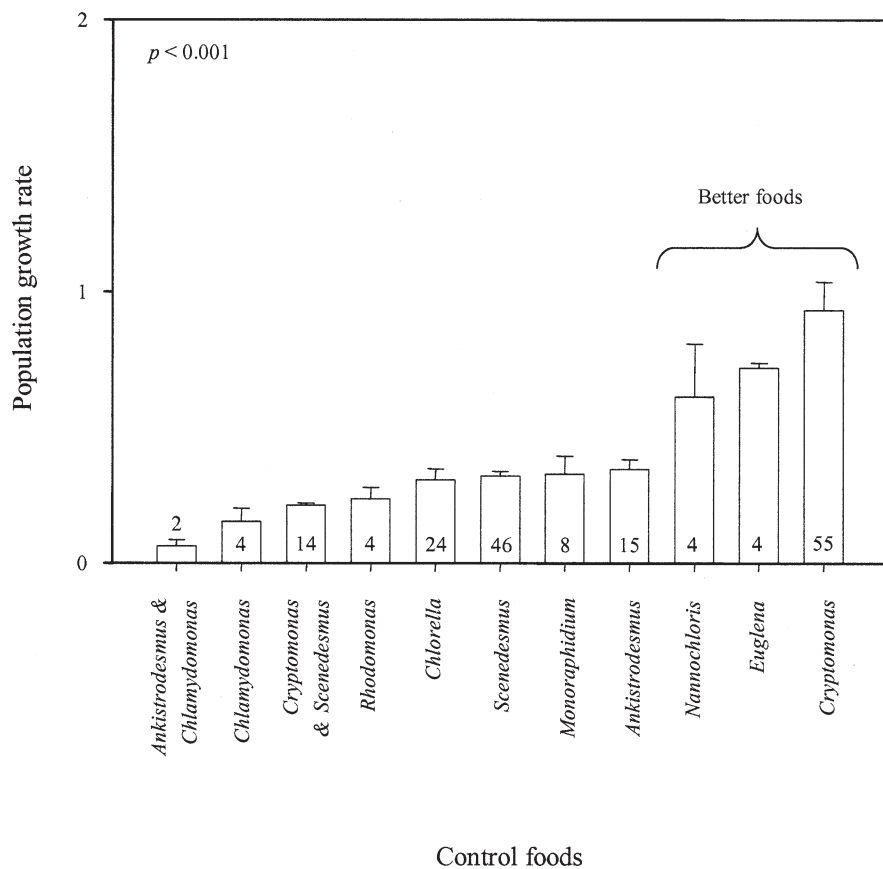


Figure A2. Control food population growth rates (mean \pm 1 SE) for grazers used in the primary meta-analysis. Inset numbers indicate sample size for each estimate where each control food growth rate estimate was used only once per study, even if multiple effect size estimates incorporated the same control food growth rate estimate more than once.

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